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## Studying the Influence of Gamma Irradiation on Germination, Vegetative Growth, Photosynthetic pigments and Biochemical Characteristics of *Dodonaea viscosa* L. Plant

Samah Mostafa El-Sayed<sup>1</sup>, Amr Said Mohamed<sup>2</sup> and Ramez Saber Thabet<sup>3</sup>

<sup>1</sup>Ornamental Plants and Woody Trees Dept., National Research Centre (NRC), Dokki, Giza, Egypt.

<sup>2</sup>Botanical Gardens Research Dept., Horticultural Research Institute (HRI), Agricultural Research Center (ARC), Giza, Egypt.

<sup>3</sup>Ornamental Plants and Landscape Gardening Research Dept. Horticultural Research Institute (HRI), Agricultural Research Center (ARC), Giza, Egypt.

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### ABSTRACT

**Background:** *Dodonaea viscosa* has a fibrous expansion root system and is effective in soil stabilization, which reduces erosion of sand dunes. It has green leaves and deep red capsules, which make it pleasing to see and can also be used as a hedge, specimen plant, or maybe a small patio tree. Gamma rays are considered a source of some desirable mutations in plants. Therefore, the goal was to study the effect of different concentrations of gamma rays to obtain the best concentration that produces the best and strongest specifications for the plant. **Results:** The results obtained through the study showed that the seeds exposed to the dose of 200 Gy had a positive effect on FGP, GRI, and CVG, while the GI and TSG were affected by the dose of 300 Gy. On the other hand, the seeds treated with the dose of 400 Gy had a significant effect on most of the vegetative characteristics, photosynthetic pigments, and the activity of PPO and POD isoenzymes. Data analysis of (SDS-PAGE) showed a change in protein metabolism, which is one of the functional aspects of the effects of radiation. **Conclusions:** The results showed that the doses of 200 and 300 Gy had a positive effect on germination characteristics, while the morphological and chemical differences appeared mostly in plants produced from seeds treated with 400 Gy.

**Keywords:** *Dodonaea viscosa*, gamma ray, vegetative growth, photosynthetic pigments, Native-PAGE, SDS-PAGE, superoxide dismutase, peroxidase, polyphenol oxidase.

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### 1. Introduction

*Dodonaea* genus belongs to the family Sapindaceae and comprises 68 species, most of which are small trees and shrubs (Muhammad *et al.*, 2016). *Dodonaea viscosa* (L.) Jacq., common name is hopbush. It is a flowering evergreen shrub, multi-stemmed shrub, or single-stemmed small tree up to 5–7 m tall (Mothana *et al.*, 2010). *Dodonaea viscosa* originated in Australia and is available throughout tropical and subtropical countries (AL-Oraimi and Hossain, 2016), *D. viscosa* has a fibrous expansion root system and is effective in soil stabilization, which reduces erosion of sand dunes. It can be tolerated during a drought (Lira-Caballero *et al.*, 2018; Ilyas *et al.*, 2020). *D. viscosa* is an aesthetically pleasing plant. It has green leaves and deep red capsules, which make it pleasing to see and can also be used as a hedge, specimen plant, or maybe a small patio tree. The wood is suitable for use as thatching to cover the roofs of houses (Al-Snafi, 2017).

More studies have shown that *D. viscosa* plants include most of the main secondary metabolites. Numerous chemical components were isolated (Mostafa *et al.*, 2014), which have numerous pharmaceutical properties and are conventionally used around the world. It is used to treat

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**Corresponding Author:** Samah Mostafa El-Sayed, Ornamental Plants and Woody Trees Dept., National Research Centre (NRC), Dokki, Giza, Egypt.  
E-mail: ensamah\_83@hotmail.com

skin diseases (Priya *et al.*, 2021), and is also used as an antimalarial (Clarkson *et al.*, 2004; Mengiste *et al.*, 2012), antidiabetic (Hossain, 2019), and antibacterial agent (Anandan *et al.*, 2019).

The gamma ray is a kind of electromagnetic radiation with a short wavelength that has a good penetration effect on molecules, causing ionization of materials through the excitement of their electrons (Majeed *et al.*, 2018). The host genetic material was disturbed in the ionized cells, which caused significant changes in the inherited traits. Cobalt-60 is one of the main sources of gamma rays used in radiobiological work at present (Moussa *et al.*, 2006). At the level of medicinal plant studies, radiation is a good inducer for many useful improvements. Good results have been obtained at the level of germination, yield growth parameters, and enhanced active ingredients (Borzouei *et al.*, 2010; Mali *et al.*, 2011; Silva *et al.*, 2013; Kumagai and Takahashi, 2020)

Gamma rays represent an effective mutagenic tool for plant breeders who want to add new traits to commercially valuable crops and develop new varieties (Wi *et al.*, 2007). Gamma-rays directly interact with the cell parts, reaching the membranes, nucleic acids, and proteins (Araujo *et al.*, 2016). However, an indirect action is also recorded through the reactive oxygen species (ROS) generation from water radiolysis (Qi *et al.*, 2015; Tanabe *et al.*, 2022). The physiological and metabolic processes are tremendously changed due to gamma radiation, which leads to the induction of antioxidants and polyphenolic compounds that act as primary defense mechanisms (Khan *et al.*, 2018).

The disturbance of DNA could be constant, which leads to serious impacts, or might be temporal as a result of the ability of DNA to restore itself after the injury following the mechanism of nucleotides in restoring themselves (Shi *et al.*, 2017; Tiwari and Wilson III, 2019).

Determination of the optimum dose of gamma rays to induce mutation in each species is very important in order to get a higher mutant frequency. Hence, the aims of this study were to determine the radio sensitivity and investigate the effects of various doses of gamma irradiation on germination traits, plant growth, morphological variation, and biochemical genetic characteristics, and discuss their potential for *Dodonaea viscosa* ornamental. Hitherto, there is no previous report of an induced mutation in this species.

## 2. Materials and Methods

### 2.1. Seeds collection and experiment location

Seeds of *Dodonaea viscosa* were obtained from the Agricultural Research Center, Giza, located at 30°01'12.5"N 31°12'24.2"E, Egypt. The field experiment was carried out during 2020/2021 season in the experimental farm of Horticulture Research Institute of Agricultural Research Center, Cairo, Egypt. The chemical analysis were conducted in the laboratory of Ornamental Plants and Woody Trees Department, National Research Centre.

### 2.2. Procedure

The uniform healthy seeds of *Dodonaea viscosa* were sterilized for 15 minutes in a 3.75% sodium hypochlorite solution (Telci *et al.*, 2011) and then Irradiation of seeds was performed using a 60Co (Cobalt 60) gamma source (Gamma Chamber 900) in ambient conditions at the National Center for Radiation Research and Technology, Nasr City, Cairo, Egypt. The doses of exposure were (0, 100, 200, 300, 400, and 500 Gray), where every treatment contains 20 seeds. The irradiated seeds were cultivated individually in March 2020 in plastic bags containing the soil mixture of clay and sand (1:1 v/v). Physical and chemical analysis were carried out according to Jackson (1973) and tabulated in Table (1), and the obtained seedlings were harvested in March 2021.

**Table 1:** The physical and chemical properties of the soil mixture

Soil sample	Coarse sand%		Fine sand%			Silt%	Clay%			
	61.44		9.36			12	17.20			
Sandy loam	E.C.(1:1) (dS/m)	pH	O.M. (%)	Anion (meq/l)			Cation (meq/l)			
				HCO <sub>3</sub> <sup>-</sup>	Cl <sup>-</sup>	SO <sub>4</sub> <sup>-</sup>	Ca <sup>++</sup>	Mg <sup>++</sup>	Na <sup>++</sup>	K <sup>+</sup>
	0.48	8.1	1.36	6.88	3.25	2.47	6.00	1.82	2.79	0.78

## 2.3. Data recorded

### 2.3.1. Germination parameters

#### Final Germination percentage (FGP)

According to Anjum and Bajwa (2005), Final Germination percentage (FGP) was calculated as follows:

$$FGP = (NT/N) \times 100$$

Where NT is the number of germinated seeds at each time to count and N is the total seeds in each treatment.

#### Germination index (GI)

The germination index is a quantitative expression of germination that relates the daily germination rate to the maximum germination value. The germination index (GI) was calculated as described by the Association of Official Seed Analysts (Dezfuli *et al.*, 2008) by the following formula:

$$GI = N1/1 + N2/2 + N3/3 + \dots + Nn/n$$

Where; N1, N2, N3, ... Nn are number of germinated seeds, (1, 2, 3, ...) are days of the count and n is day of final count

#### Germination Rate Index (GRI)

The Germination Rate Index (GRI) was determined as described by Esechi, 1994:

$$GRI (\% / \text{day}) = G1/1 + G2/2 + \dots + Gx/x,$$

Where; G1 is germination percentage at the first day after sowing and G2 is germination percentage at the second day after sowing.

#### Time Spread of Germination (TSG)

TSG (day) is the time in days between the first and last germination occurring in a seed part, was described by Kader (1998)

#### Coefficient of Velocity of Germination (CVG)

The Coefficient of Velocity of Germination (CVG) Jones and Sanders (1987) gives an indication of the rapidity of germination. Its value increases when the number of germinated seeds increases and the time required for germination decreases (Talská *et al.*, 2020).

$$CVG = N1 + N2 + \dots + Nx/100 \times N1T1 + \dots + NxTx$$

Where; N is No. of seeds germinated each day and T is No. of days from seeding corresponding to germination

#### Mean germination time (MGT).

The Mean Germination Time (MGT) (Mavi *et al.*, 2010) computes the day of average germination; accordingly, the lower the MGT, the faster a population of seeds reaches germination.

$$MGT (\text{day}) = \sum NiTi / \sum Ni$$

Where Ni is number of seeds used in the treatment and Ti represents the number of days since the beginning of the experiment.

### 2.3.2. Vegetative growth parameters

Plant height (cm), root length (cm), stem diameter (cm), leaf area (cm<sup>2</sup>), number of leaves/plant and number of branches/plant.

### 2.3.3. Photosynthetic pigments

Photosynthetic pigments including chlorophyll a, b, and carotenoids (mg. g<sup>-1</sup> F.W.) were

determined according to Saric *et al.*, (1967).

### 2.3.4. Gene expression (POD, PPO, and SDS-PAGE)

#### Native Polyacrylamide Gel Electrophoresis (Native-PAGE)

Electrophoresis was performed to identify isoenzyme differences between control and treatments in the second season.

Peroxidase isoenzymes POD (E.C. 1.11.1.7) in leaf samples were assessed by the procedure defined by (Barceló *et al.*, 1987).

Polyphenol oxidase isoenzymes PPO (E.C. 1.10.3.1) in leaves (100 mg fresh weight) samples were estimated as described by Thipyapong *et al.*, (1995).

The relative distance (Rf-value) of the bands on the gel was calculated as described by (Manganaris and Alston, 1992) using  $rf = 1.0$ , distance to the fastest band, and  $Rf = 0.0$ , the starting point.

#### Sodium Dodecyl Sulfate Polyacrylamide Gel Electrophoresis (SDS-PAGE)

*Dodonaea viscosa* leaf protein fingerprints were analyzed using SDS-PAGE Sodium Dodecyl Sulfate-Polyacrylamide Gel Electrophoresis (SDS-PAGE) based on the method of (Laemmli, 1970) as modified by (Studier, 1973). The molecular weight of proteins was then obtained relative to the marker, a large variety of molecular weight proteins (Gene Direx com).

### 2.4. Experimental layout and statistical analysis

The experimental layout was set in randomized complete design (RCD). The data were analyzed through analysis of variance ANOVA and the treatments' means were compared for significance by Least Significant Difference (LSD) test at the 5% level as described by Little and Hills (1978). Standard deviation ( $\pm$ SD) was calculated.

## 3. Results

### 3.1. Germination parameters

Gamma rays treatment at doses 0, 100, 200, 300, 400, and 500 Gy showed different effects on the germination parameters of *D. viscosa* seeds, where FGP, GRI, and CVG were raised in the seeds exposed to dose 200 Gy giving values  $95 \pm 2.65\%$ ,  $6.81 \pm 0.68$  and  $6.46 \pm 0.89$ , respectively, while at the same dose MGT day gave the lowest value  $15.44 \pm 1.31$  when compared with untreated seeds. GI and TSG gave the highest value in the seeds exposed to 300 Gy as compared with untreated seeds.

**Table 2:** The effect of gamma rays treatment with different doses on germination parameters of *D. viscosa* seeds

Dose	FGP (%)	GI	TSG (day)	GRI (%/ day)	CVG	MGT (day)
Control (0.0)	80 $\pm$ 2.65	996 $\pm$ 4.58	21 $\pm$ 2.00	4.45 $\pm$ 0.53	5.06 $\pm$ 0.35	19.75 $\pm$ 2.13
100Gy	82 $\pm$ 3.00	1386 $\pm$ 8.19	24 $\pm$ 2.00	5.7 $\pm$ 1.08	5.85 $\pm$ 0.54	17.09 $\pm$ 1.67
200Gy	95 $\pm$ 2.65	1573 $\pm$ 7.55	22 $\pm$ 2.65	6.81 $\pm$ 0.68	6.46 $\pm$ 0.89	15.44 $\pm$ 1.31
300Gy	93 $\pm$ 2.65	1692 $\pm$ 10.44	27 $\pm$ 1.00	6.43 $\pm$ 0.95	5.95 $\pm$ 1.02	16.81 $\pm$ 2.01
400Gy	90 $\pm$ 2.65	1535 $\pm$ 10.44	24 $\pm$ 4.36	5.91 $\pm$ 0.94	3.72 $\pm$ 0.86	17.94 $\pm$ 1.44
500Gy	85 $\pm$ 4.36	1437 $\pm$ 7.81	25 $\pm$ 4.36	5.33 $\pm$ 0.50	5.53 $\pm$ 0.73	18.09 $\pm$ 1.16
LSD <sub>at 5%</sub>	<b>5.43</b>	<b>18.37</b>	<b>5.34</b>	<b>1.48</b>	<b>1.36</b>	<b>2.96</b>

### 3.2. Vegetative growth parameters

The obtained results in Table (3) and Fig. (1) Showed that the effect of different radiation doses was evident on plants, as it showed significant differences in all morphological characteristics. It was clear that the radiation treatment at 400 Gy gave the highest values in the following traits: plant length (cm), number of leaves per plant, number of branches per plant, leaf area (cm<sup>2</sup>), and stem diameter (cm), which were (41.62, 53.33, 9.33, 6.52 and 0.38, respectively), while the highest value was in the root length when seeds were treated with 200 Gy (28.69 cm) when compared to the non-irradiated plants.

### 3.3. Photosynthetic pigments

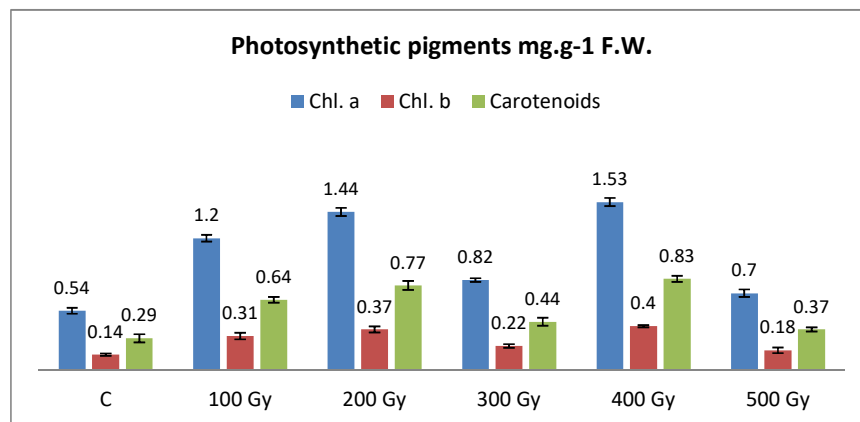
The data presented in Fig. (2) Stated that all gamma rays treatments increased the content of the photosynthetic pigments. Where; the seeds were exposed to gamma rays at 400Gy produced the highest content of chlorophyll a, b and carotenoids giving values 1.53, 0.40 and 0.83 mg.g<sup>-1</sup> F.W., respectively, followed by treatment 200Gy which gave 1.44, 0.37 and 0.77 mg.g<sup>-1</sup> F.W., respectively, as compared to Non-Irradiated Seeds.



**Fig. 1:** The differences between morphological traits of irradiated *D. viscosa* with gamma rays (Gy) where; C: control (0.0), 1:100, 2: 200, 3: 300, 4:400 and 5: 500.

**Table 3:** The effect of gamma rays treatment with different doses on the vegetative growth traits of *D. viscosa* seedlings

Dose	Plant height (cm)	No. of Leaves/ plant	No. of branches/ plant	Leaf area (cm <sup>2</sup> )	Stem diameter (cm)	Root length (cm)
control	22.5 ± 1.80	28.33±2.35	1.00±0.00	2.05±0.09	0.22±0.03	20.80±1.61
100Gy	33.52 ± 2.54	36.67±2.40	5.33±1.15	4.28±0.10	0.30±0.04	23.75±2.91
200Gy	34.27 ± 1.68	48.00±2.21	7.00±1.00	4.50±0.26	0.34±0.04	28.69±1.57
300Gy	30.57 ± 2.81	35.67±2.39	4.67±1.15	3.33±0.30	0.28±0.05	22.43±1.99
400Gy	41.62 ± 2.42	53.33±2.81	9.33±2.08	6.25±0.08	0.38±0.04	26.49±3.37
500Gy	28.50 ± 1.56	30.33±2.58	2.67±0.58	2.92±0.10	0.27±0.04	21.53±1.63
LSD at 5%	3.89	4.38	2.09	0.32	0.072	4.07



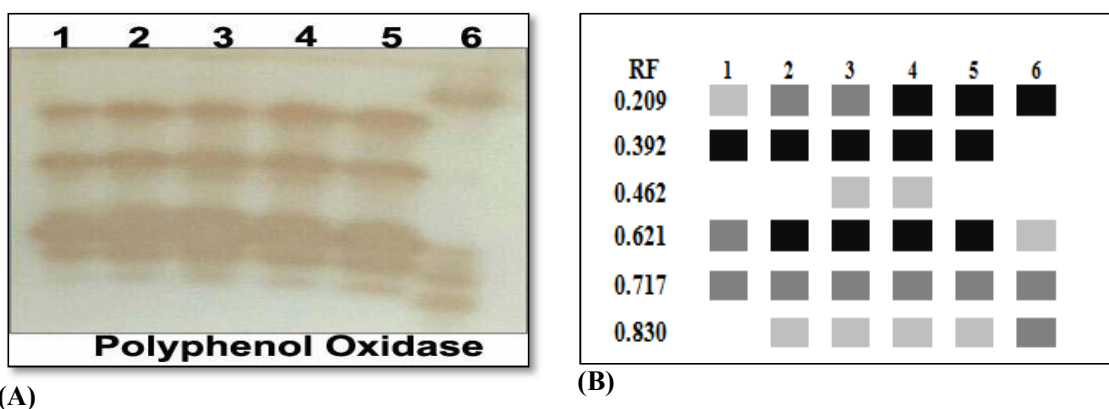
**Fig. 2:** The effect of gamma rays treatment with different doses on the photosynthetic pigments (chlorophyll a, b and carotenoids mg.g<sup>-1</sup> F.W.) of *D. viscosa* seedlings

### 3.4. Gene expression

#### 3.4.1. Native Polyacrylamide Gel Electrophoresis (Native-PAGE Antioxidant Isoenzyme

##### 3.4.1.1. Polyphenol Oxidase isoenzymes

The electrophoretic pattern and diagram of polyphenol oxidase isoenzymes are illustrated in Fig. (3 A&B) and Table (4), showed the activity stain for isoenzymes, gamma irradiation effects appeared on both band numbers and intensity (faint, moderate and high intensities) when compared with non-irradiation plant. The PPO isozyme total bands of *Dodonaea viscosa* a plant leaves showed six PPO isoenzymes at Rf (0.209, 0.392, 0.462, 0.621, 0.717 and 0.830). The effect of gamma irradiation on number of bands was clear at both 200 and 300Gy, which gave the highest value of band numbers (6 bands), and that the third and sixth bands at Rf (0.462 and 0.830) are positive markers at the 200 and 300Gy doses compared with untreated plant which gave 4 bands. The intensity of the PPO activity bands at 300 and 400 Gy produced three highly intense bands at Rf (0.209, 0.392 and 0.621), while a control got only one highly intense band Rf at (0.392). The activity (number and intensity bands) of the PPO increased with increased gamma irradiation up to 400 Gy then decreased at 500 Gy.



**Fig. 3:** Effect of different doses of gamma rays (Gy) where; 1: Control (0) , 2: 100, 3: 200, 4: 300, 5: 400, 6: 500 (A) polyphenol oxidase isoenzyme and (B) ideogram analysis of polyphenol oxidase isoenzyme of *D. viscosa* plants

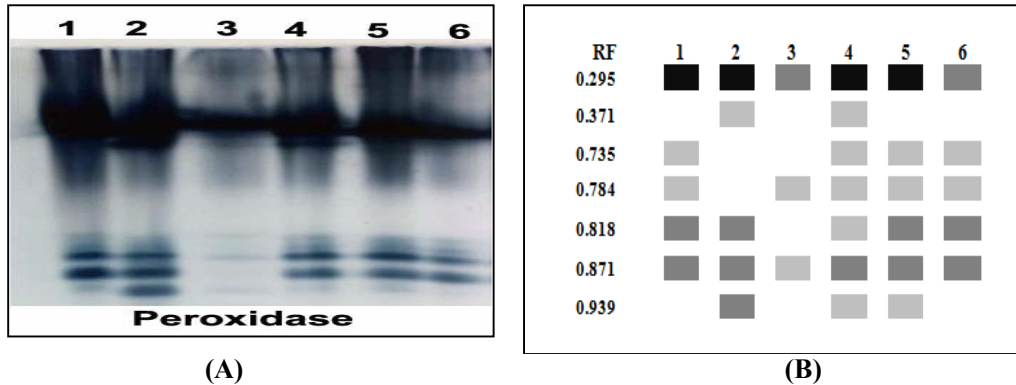
**Table 4:** Isomers of polyphenol oxidase isoenzymes (+/-) and their retention factor (Rf) in response to different doses of gamma rays (Gy) Scheme, where; 1: Contro (0) , 2: 100, 3:200, 4: 300, 5: 400, 6: 500

RF	1	2	3	4	5	6
0.209	+	++	++	+++	+++	+++
0.392	+++	+++	+++	+++	+++	—
0.462	—	—	+	+	—	—
0.621	++	+++	+++	+++	+++	+
0.717	++	++	++	++	++	++
0.830	—	+	+	+	+	++

##### 3.4.1.2. Peroxidase isoenzyme

The influence of different gamma rays doses on the activity of Peroxidase isoenzyme of *Dodonaea viscosa* plants is shown in Fig. (4 A & B) and Table (5) The POD isozyme total bands of *D.viscosa* plant leaves inducted 7 POD isozymes at Rf (0.295, 0.371, 0.735, 0.784, 0.818, 0.871, and 0.939). The highly POD activity of gamma irradiation based on number of bands was clear at 300 gy, which gave the highest number of bands (7 bands), then treatment with gamma irradiation at 400 Gy, which induced 6 bands, while the control showed 5 bands. The highly of the POD activity bands recorded at 100 Gy gave 1 highly intense band at Rf (0.295) and three moderate intensity bands at Rf (0.818, 0.871, and 0.939) while a control recorded 1 highly intense band at Rf (0.295), two moderate bands at Rf (0.818 and 0.871), and two faint bands at Rf (0.735 and 0.784). The lowest intensity

bands were at irradiation at 200 Gy, which produced three bands one moderate band at Rf (0.295) and two faint bands at Rf (0.784 and 0.871).



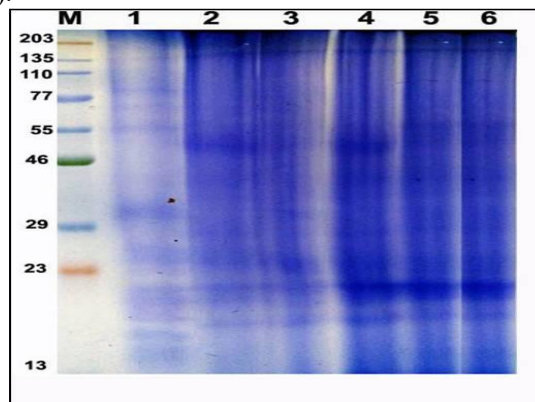
**Fig. 4:** Effect of different doses of gamma rays (Gy) where; 1:Contro (0) , 2: 100, 3: 200, 4: 300, 5: 400Gy, 6: 500 (A) peroxidase isoenzyme and (B) ideogram analysis of Peroxidase isoenzyme of *D. viscosa* plants

**Table 5:** Isomers of peroxidase isoenzymes (+/-) and their retention factor (Rf) in response to different doses of gamma rays (Gy) Scheme, where; 1:control (0) , 2: 100, 3:200, 4: 300, 5: 400, 6: 500

RF	1	2	3	4	5	6
0.295	+++	+++	—	+++	+++	++
0.371	—	+	—	+	—	—
0.735	+	—	—	+	+	+
0.784	+	—	+	+	+	+
0.818	++	++	—	+	++	++
0.871	++	++	+	++	++	++
0.939	—	++	—	+	+	—

### 3.4.1.3. Sodium Dodecyl Sulfate Polyacrylamide Gel Electrophoresis (SDS-PAGE)S protein banding pattern

As shown in Fig. (5) and Table (6), The total proteins bands from the leaves generated from IS (Irradiated Seeds) and NIS (Non-Irradiated Seeds) were analyzed by SDS-PAGE in the range of 13.005-131.906 kDa, four monomorphic bands (Similar protein bands in all treatments) were observed with molecular weights (14.849, 17.689, 19.143 and 131.906 kDa), only 2 unique bands were different between the control and the treated sample showed up in (0.0 gy) with molecular weights (35.7 and 25.8 kDa). A 101.173 kDa protein bands was present in (0.0, 400 and 500 gy) and absent in the rest of the treatments, also protein band of 70.701 kDa was present only with treatments (0.0, 300, 400 and 500 gy).



**Fig. 5:** Protein banding pattern between 1: control (0.0), 2: (100), 3: (200), 4: (300), 500 (400) and 6: (500) gy *Dodonia viscosa* protein banding pattern by SDS-PAGE analysis.

Protein band at 61.571 does not appear in control (0.0 gy) but appears in (100,300, 400 and 500 gy). It is observed that four bands were monomorphic, while six bands were polymorphic and unique, giving 60% polymorphism. This is due to the irradiation effect on the protein profile.

**Table 6:** Gamma rays doses (Gy) (1):control (0.0), (2):100, (3): 200, (4): 300, (5):400 and (6): 500 *Dodonia viscosa* protein banding pattern by SDS-PAGE analysis.

MW	1	2	3	4	5	6	Polymorphism
131.906	1	1	1	1	1	1	Monomorphic
101.173	1	0	0	0	1	1	Polymorphic
70.701	1	0	0	1	1	1	Polymorphic
61.571	0	1	0	1	1	1	Polymorphic
35.715	1	0	0	0	0	0	Unique
25.745	1	0	0	0	0	0	Unique
19.143	1	1	1	1	1	1	Monomorphic
17.689	1	1	1	1	1	1	Monomorphic
14.849	1	1	1	1	1	1	Monomorphic
13.005	1	0	0	0	1	1	Polymorphic

#### 4. Discussion

It was observed that the germination percentage increased from the beginning of the radiation doses, where the highest percentage was at 200Gy and then decreased gradually again with increasing the dose up to 500Gy. These findings positively agree with germination traits studies in *Lathyrus chrysanthus* (Beyaz *et al.*, 2016), *Ocimum basilicum* (Asgari Lajayer *et al.*, 2019), and *Pavonia* sp. (Yue and Ruter, 2020). The stimulating effects of gamma rays on germination may be attributed to the activation of RNA or protein synthesis, which occurred during the early stage of germination after seeds were irradiated (Abdel-Hady *et al.*, 2008); (Araujo *et al.*, 2016) or may be attributed to the effects of radiation on genes controlling these traits, stimulating hormones, activating enzymes involved in germination processes, and accelerating DNA repair. In addition, it may improve plant germination by accelerating cell division in meristematic tissues (Dhakshanamoorthy *et al.*, 2011); (Katiyar *et al.*, 2022).

Exposure to high doses led to a decrease in germination traits, this decrement might be due to the effect of mutagens on meristematic tissues of the seed as well as chromosomal aberrations and interruptions in DNA replication and growth regulators (Layek *et al.*, 2022; Wang *et al.*, 2017). On the other hand, the inhibition of seed germination at high doses could be due to the damage to seed tissue, chromosomes, and subsequent mitotic retardation, and the severity of the damage depends on the doses used (Li *et al.*, 2018); (Volkova *et al.*, 2020).

The results obtained through this study showed different effects of the gamma ray doses used on the morphological characteristics of *D. viscosa* seedlings, with the highest values of the traits under study obtained from treatment with the 400Gy dose. The increased plant growth caused by gamma irradiation treatment could be explained by the stimulation of the biosynthesis of some amino acids (Hanafy and Akladious, 2018), acceleration in cell division rates (Majeed *et al.*, 2018) as well as activation of auxin (Qi *et al.*, 2015). The stimulatory effect on the morphological traits is in line with Beyaz *et al.* (2016) on *Lathyrus chrysanthus*, (Ryu *et al.*, 2019) on *Chrysanthemum morifolium*, Altay *et al.* (2019) on *Ocimum basilicum* and Cahyaningsih *et al.* (2022) on *Echinacea purpurea*.

The results revealed that gamma rays had a positive effect on the content of photosynthetic pigments, as the highest content appeared when the treatment was 400gy. El-Beltagi *et al.* (2022) attributed the stimulating effect of irradiation on chlorophyll to stabilizing the enzyme active site and photosynthetic reactions. The highly effective effects of gamma radiation on chemical content may be induced by irradiation induced stimulation of plant growth at low doses, particularly during the early genetic stages, which may be regulated through changes in plant growth hormones. The effect of radiation on increasing endogenous hormones in plants and the involvement of auxin and cytokinin in regulating growth and photosynthetic attributes are known (Xiong *et al.*, 2020). The present results are generally in agreement with those of (Patil *et al.*, 2017) on *Dendrathera grandiflora*; (Wang *et*



*al.*, 2020) on *Chrysanthemum morifolium* ‘Donglinruixue;Asgari Lajayer *et al.*, (2019) on *Ocimum basilicum*, Hajizadeh *et al.* (2022) on *Lilium longiflorum*; and Ghosh *et al.* (2020) on *J. auriculatum*.

The resulted PPO expression activity increased with increasing radiation dose up to 400 gy compared with the control (0.0) Gy, in agreement with Mohamed *et al.*, (2021),while POD activity declined at 200 gy and increased at 100, 300, and 400 gy, in agreement with Li *et al.*, (2022) they said that gamma radiation is a stress that leads to the creation of reactive oxygen species (ROS) such as superoxides, hydrogen peroxide, and hydroxyl radicals, which are likely to cause oxidative stress. Increased enzyme activity is primarily responsible for the plant’s improved antioxidant capacity and activity following irradiation (Bitarishvili *et al.*, 2018).

It has been shown that gamma-radiation generates active species such as hydrated electrons and hydroxyl radicals (OH.) through its impact on water molecules, which then react with protein molecules (Hassan *et al.*, 2018).The appearance and disappearance of protein bands may be associated with radiation that affect gene expression alterations agreement with (Singh and Datta, 2010) that used a Cobalt-60 gamma source to irradiate wheat seeds and observed an increase in qualitative changes in the protein profiles of the irradiated seeds. Irradiation-induced chemical changes in proteins include degradation, cross-linking, disturbance of the ordered structure of protein molecules, and aggregation of polypeptide chains produced by oxygen radicals Ariraman *et al.*, (2016).

## 5. Conclusions

Treatment with gamma rays at different concentrations had positive effects on germination, vegetative characteristics, photosynthetic pigments in leaves, enzymatic activity of antioxidant enzymes and plant protein content. Seeds exposed to the dose of 200 Gy showed positive effects on some germination characteristics, while others were affected by the dose of 300 Gy. Whereas, the vegetative characteristics, photosynthetic pigments and enzymatic activity were positively affected by the dose of 400 Gy.

## List of Abbreviations

*D. viscosa*: *Dodonaea viscosa*, FGP: Final Germination percentage, GI: Germination index, GRI: Germination Rate Index, CVG: Coefficient of Velocity of Germination, MGT: Mean Germination Time SDS-PAGE: Sodium Dodecyl Sulfate Polyacrylamide Gel Electrophoresis, Native-PAGE: Native Polyacrylamide Gel Electrophoresis, PPO: Polyphenol Oxidase POD: Peroxidase, SOD: Superoxide Dismutase.

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## Authors' contributions

Amr Said Mohamed carried out the field experiment and recorded the morphological data. Samah Mostafa El-Sayed performed the chemical analysis for obtained samples. Ramez Saber Thabet analyzed the obtained data statistically and wrote the manuscript. All authors read and approved the final manuscript.

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## References

Abdel-Hady, M., E. Okasha, S. Soliman, and M. Talaat, 2008. Effect of gamma radiation and gibberellic acid on germination and alkaloid production in *Atropa belladonna* L. Australian Journal of Basic and Applied Sciences, 2:401-405.

- AL-Oraimi, A.A., and M.A. Hossain, 2016. In vitro total flavonoids content and antimicrobial capacity of different organic crude extracts of *Dodonaea viscosa*. *Journal of Biologically Active Products from Nature*, 6:150-165. DOI: [10.1080/22311866.2016.1188725](https://doi.org/10.1080/22311866.2016.1188725)
- Al-Snafi, A.E., 2017. A review on *Dodonaea viscosa*: A potential medicinal plant. *IOSR Journal of Pharmacy*, 7:10-21. DOI: [10.9790/3013-0702011021](https://doi.org/10.9790/3013-0702011021)
- Altay, K., S.N. Dirim, and A.A. Hayaloglu, 2019. The effect of gamma irradiation on microbial load of purple basil (*Ocimum bacilicum* L.) leaves dried in different methods. *Journal of Food Safety*, 39:e12610. <https://doi.org/10.1111/jfs.12610>
- Anandan, M., G. Poorani, P. Boomi, K. Varunkumar, K. Anand, A.A. Chaturgoon, M. Saravanan, and H.G. Prabu, 2019. Green synthesis of anisotropic silver nanoparticles from the aqueous leaf extract of *Dodonaea viscosa* with their antibacterial and anticancer activities. *Process Biochemistry*, 80:80-88. <https://doi.org/10.1016/j.procbio.2019.02.014>
- Anjum, T., and R. Bajwa, 2005. Importance of germination indices in interpretation of allelochemical effects. *International Journal of Agriculture and Biology* 7:1560-8530.
- Araujo S.d.S., Paparella S., Dondi D., Bentivoglio A., Carbonera D., Balestrazzi A. (2016) Physical methods for seed invigoration: advantages and challenges in seed technology. *Frontiers in Plant Science*, 7:646. <https://doi.org/10.3389/fpls.2016.00646>
- Ariraman, M., T. Bharathi, and D. Dhanavel 2016. Studies on the effects of mutagens on cytotoxicity behaviour in Pigeon pea (*Cajanus cajan* (L.) Millsp) Var. CO-7. *Journal of Applied and Advanced Research*, 1:25-28. DOI: [10.21839/jaar.2016.v1i1.10](https://doi.org/10.21839/jaar.2016.v1i1.10).
- Asgari, L.B., N. Najafi, E. Moghiseh, M. Mosafieri, and J. Hadian, 2019. Effects of gamma irradiated and non-irradiated sewage sludge on growth characteristics, leaf chlorophyll index, and macronutrients concentrations in basil. *Journal of Soil Science and Plant Nutrition*, 19:580-591. DOI: [10.1007/s42729-019-00057-4](https://doi.org/10.1007/s42729-019-00057-4).
- Barceló, A.R., R. Muñoz, and F. Sabater, 1987. Lupin peroxidases. I. Isolation and characterization of cell wall-bound isoperoxidase activity. *Physiologia plantarum*, 71:448-454. <https://doi.org/10.1111/j.1399-3054.1987.tb02882.x>.
- Beyaz, R., C.T. Kahramanogullari, C. Yildiz, E.S. Darcin, and M. Yildiz, 2016. The effect of gamma radiation on seed germination and seedling growth of *Lathyrus chrysanthus* Boiss. under *in vitro* conditions. *Journal of Environmental Radioactivity*, 162:129-133. DOI: [10.1016/j.jenvrad.2016.05.006](https://doi.org/10.1016/j.jenvrad.2016.05.006).
- Bitarishvili, S., P.Y. Volkova, and S. Geras'kin, 2018.  $\gamma$ -Irradiation of barley seeds and its effect on the phytohormonal status of seedlings. *Russian Journal of Plant Physiology*, 65:446-454. <https://doi.org/10.1134/S1021443718020024>.
- Borzouei, A., M. Kafi, H. Khazaei, B. Naseriyan, and A. Majdabadi, 2010. Effects of gamma radiation on germination and physiological aspects of wheat (*Triticum aestivum* L.) seedlings. *Pak. J. Bot.*, 42:2281-2290.
- Cahyaningsih, A.P., N. Etikawati, and A. Yunus, 2022. Morphological characters variation of Indonesian accession *Echinacea purpurea* in response to gamma-ray irradiation. *Biodiversitas Journal of Biological Diversity*, 23: 5351-5359. DOI: [10.13057/biodiv/d231045](https://doi.org/10.13057/biodiv/d231045).
- Clarkson, C., V.J. Maharaj, N.R. Crouch, O.M. Grace, P. Pillay, M.G. Matsabisa, N. Bhagwandin, P.J. Smith, and P.I. Folb, 2004. In vitro antiplasmodial activity of medicinal plants native to or naturalised in South Africa. *Journal of ethnopharmacology*, 92:177-191. DOI: [10.1016/j.jep.2004.02.011](https://doi.org/10.1016/j.jep.2004.02.011)
- Dezfuli, P.M., F. Sharif-Zadeh, and M. Janmohammadi, 2008. Influence of priming techniques on seed germination behavior of maize inbred lines (*Zea mays* L.). *ARPN Journal of Agricultural and Biological Science*, 3:22-25.
- Dhakshnamoorthy, D., R. Selvaraj, and A. Chidambaram, 2011. Induced mutagenesis in *Jatropha curcas* L. using gamma rays and detection of DNA polymorphism through RAPD marker. *Comptes Rendus Biologies*, 334:24-30. <https://doi.org/10.1016/j.crvi.2010.11.004>
- El-Beltagi, H.S., R.W. Maraie, T.A. Shalaby, and A.A. Aly, 2022. Metabolites, nutritional quality and antioxidant activity of red radish roots affected by gamma rays. *Agronomy*, 12:1916. <https://doi.org/10.3390/agronomy12081916>.

- Esechie, H., 1994. Interaction of salinity and temperature on the germination of sorghum. *Journal of Agronomy and Crop Science*, 172: 194–199.
- Ghosh, S., M. Ganga, K. Soorianathasundaram, A. Kumar, and M. Kapoor, 2020. Induction of mutation in *Jasminum grandiflorum* with gamma rays and EMS and identification of novel mutants using molecular markers and SEM imaging. *Indian Journal of Horticulture*, 77:695-703. DOI: [10.5958/0974-0112.2020.00101.2](https://doi.org/10.5958/0974-0112.2020.00101.2).
- Hajizadeh, H.S., S.N. Mortazavi, F. Tohidi, H.Y.M. Helvaci, T. Alas, and V. Okatan, 2022. Effect of mutation induced by gamma-irradiation in ornamental plant liliun (*Lilium Longiflorum* Cv. Tresor). *Pak. J. Bot.*, 54:223-230. DOI: [10.30848/PJB2022-1\(23\)](https://doi.org/10.30848/PJB2022-1(23))
- Hanafy, R.S., and S.A. Akladios, 2018. Physiological and molecular studies on the effect of gamma radiation in fenugreek (*Trigonella foenum-graecum* L.) plants. *Journal of Genetic Engineering and Biotechnology*, 16:683-692. DOI: [10.1016/j.jgeb.2018.02.012](https://doi.org/10.1016/j.jgeb.2018.02.012)
- Hassan, A.B., N.S. Mahmoud, K. Elmamoun, O.Q. Adiamo, and I.A.M. Ahmed, 2018. Effects of gamma irradiation on the protein characteristics and functional properties of sesame (*Sesamum indicum* L.) seeds. *Radiation Physics and Chemistry*, 144:85-91. <https://doi.org/10.1016/j.radphyschem.2017.11.020>
- Hossain, M.A., 2019. Biological and phytochemicals review of Omani medicinal plant *Dodonaea viscosa*. *Journal of King Saud University-Science*, 31:1089-1094. <https://doi.org/10.1016/j.jksus.2018.09.012>
- Ilyas, N., R. Mazhar, H. Yasmin, W. Khan, S. Iqbal, H.E. Enshasy, and D.J. Dailin, 2020. Rhizobacteria isolated from saline soil induce systemic tolerance in wheat (*Triticum aestivum* L.) against salinity stress. *Agronomy*, 10:989. <https://doi.org/10.3390/agronomy10070989>.
- Jackson, M.L. (1973). *Soil Chemical Analysis*. Prentice-Hall, Inc. Limited, New York. p. 125-179.
- Jones, K.W. and D. Sanders, 1987. The influence of soaking pepper seed in water or potassium salt solutions on germination at three temperatures. *Journal of Seed Technology*, 97-102. <http://www.jstor.org/stable/23432941>
- Kader (Al-Mudaris), M., 1998. Notes on various parameters recording the speed of seed germination. *Journal of Agriculture in the Tropics and Subtropics*, 99, 147–154.
- Katiyar, P., N. Pandey, and S. Keshavkant, 2022. Gamma Radiation: A Potential Tool for Abiotic Stress Mitigation and Management of Agroecosystem. *Plant Stress*:100089. <https://doi.org/10.1016/j.stress.2022.100089>.
- Khan, M.I., J.H. Shin, and J.D. Kim, 2018. The promising future of microalgae: current status, challenges, and optimization of a sustainable and renewable industry for biofuels, feed, and other products. *Microbial cell factories*, 17:1-21. <https://doi.org/10.1186/s12934-018-0879-x>.
- Kumagai, E., and T. Takahashi, 2020. Soybean (*Glycine max* (L.) Merr.) Yield reduction due to late sowing as a function of radiation interception and use in a cool region of Northern Japan. *Agronomy*, 10:66. DOI: [10.3390/agronomy10010066](https://doi.org/10.3390/agronomy10010066)
- Laemmli, U.K., 1970. Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature*, 227:680-685. <https://doi.org/10.1038/227680a0>
- Layek, S., S. Pramanik, A. Das, A.K. Gupta, A. Bhunia, and M.K. Pandit, 2022. Effect of gamma radiation on seed germination and seedling growth of snake gourd (*Trichosanthes anguina* L.). *South African Journal of Botany*, 145:320-322. <https://doi.org/10.1016/j.sajb.2021.07.039>.
- Li, M., L. You, J. Xue, and Y. Lu, 2018. Ionizing radiation-induced cellular senescence in normal, non-transformed cells and the involved DNA damage response: a mini review. *Frontiers in pharmacology*, 9:522. DOI: [10.3389/fphar.2018.00522](https://doi.org/10.3389/fphar.2018.00522)
- Li, Y., L. Chen, X. Zhan, L. Liu, F. Feng, Z. Guo, D. Wang, and H. Chen, 2022. Biological effects of gamma-ray radiation on tulip (*Tulipa gesneriana* L.). *Peer J.*, 10:e12792. <https://doi.org/10.7717/peerj.12792>
- Lira-Caballero, V.G., M.R. Martínez-Menez, A. Romero-Manzanares, and E. García-Moya, 2018. Initial floristic composition of rehabilitated gullies through bioengineering in the Mixteca Region, Sierra Madre del Sur, Mexico. *Journal of Mountain Science*, 15:2120-2135. <https://doi.org/10.1007/s11629-018-4899-0>

- Little, T. M. and Hills, F. J., 1978. Agricultural Experimentation - Design and Analysis. John Wiley and Sons, Inc., pp. 53-60.
- Majeed, A., Z. Muhammad, R. Ullah, and H. Ali, 2018. Gamma irradiation i: effect on germination and general growth characteristics of plants—a review. *Pakistan Journal of Botany*, 50:2449-2453.
- Mali, A.B., K. Khedkar, and S.S. Lele, 2011. Effect of gamma irradiation on total phenolic content and in vitro antioxidant activity of pomegranate (*Punica granatum* L.) peels. *Food and Nutrition Sciences*, 2:428. DOI: [10.4236/fns.2011.25060](https://doi.org/10.4236/fns.2011.25060)
- Manganaris, A., and F. Alston, 1992. Inheritance and linkage relationships of peroxidase isoenzymes in apple. *Theoretical and Applied Genetics*, 83:392-399. <https://doi.org/10.1007/BF00224288>.
- Mavi, K., I. Demir, and S. Matthews, 2010. Mean germination time estimates the relative emergence of seed lots of three cucurbit crops under stress conditions. *Seed Science and Technology*, 38:14-25. DOI: [10.15258/sst.2010.38.1.02](https://doi.org/10.15258/sst.2010.38.1.02)
- Mengiste, B., E. Makonnen, and K. Uрга, 2012. *In vivo* antimalarial activity of *Dodonaea Angustifolia* seed extracts against *Plasmodium berghei* in mice model. *Momona Ethiopian Journal of Science*, 4:47-63. DOI: [10.4314/mejs.v4i1.74056](https://doi.org/10.4314/mejs.v4i1.74056)
- Mohamed, E.A., E. Osama, E. Manal, A. Samah, G. Salah, K.M. Hazem, W. Jacek, and E. Nabil , 2021. Impact of gamma irradiation pretreatment on biochemical and molecular responses of potato growing under salt stress. *Chemical and Biological Technologies in Agriculture*, 8:1-11. <https://doi.org/10.1186/s40538-021-00233-8>
- Mostafa, A.E., A. Atef, A.-E.I. Mohammad, M. Jacob, S.J. Cutler, and S.A. Ross, 2014. New secondary metabolites from *Dodonaea viscosa*. *Phytochemistry Letters*, 8:10-15. <https://doi.org/10.1016/j.phytol.2013.12.008>
- Mothana, R.A., S.A. Abdo, S. Hasson, F. Althawab, S.A. Alaghbari, and U. Lindequist, 2010. Antimicrobial, antioxidant and cytotoxic activities and phytochemical screening of some yemeni medicinal plants. *Evidence-based Complementary and alternative medicine*, 7:323-330. doi: [10.1093/ecam/nen004](https://doi.org/10.1093/ecam/nen004)
- Moussa, M., D. Sumanasekera, S. Ibrahim, H. Lubberding, C. Hooijmans, H. Gijzen, and M. Van Loosdrecht, 2006. Long term effects of salt on activity, population structure and flocc characteristics in enriched bacterial cultures of nitrifiers. *Water research*, 40:1377-1388. DOI: [10.1016/j.watres.2006.01.029](https://doi.org/10.1016/j.watres.2006.01.029)
- Muhammad, A., G. Tel-Çayan, M. Öztürk, M.E. Duru, S. Nadeem, I. Anis, S.W. Ng, and M.R. Shah, 2016. Phytochemicals from *Dodonaea viscosa* and their antioxidant and anticholinesterase activities with structure–activity relationships. *Pharmaceutical Biology*, 54:1649-1655. DOI: [10.3109/13880209.2015.1113992](https://doi.org/10.3109/13880209.2015.1113992)
- Patil, U., A. Karale, S. Katwate, and M. Patil, 2017. Mutation breeding in chrysanthemum (*Dendrathera grandiflora* T.). *Journal of Pharmacognosy and Phytochemistry*, 6:230-232.
- Priya, V.T., N. Balasubramanian, V. Shanmugaiah, P. Sathishkumar, N. Kannan, C. Karunakaran, A. Alfarhan, and P. Antonisamy, 2021. Partially purified lead molecules from *Dodonaea viscosa* and their antimicrobial efficacy against infectious human pathogens. *Journal of Infection and Public Health*, 14:1822-1830. <https://doi.org/10.1016/j.jiph.2021.11.007>
- Qi, W., L. Zhang, W. Feng, H. Xu, L. Wang, and Z. Jiao, 2015. ROS and ABA signaling are involved in the growth stimulation induced by low-dose gamma irradiation in *Arabidopsis* seedling. *Applied biochemistry and biotechnology*, 175:1490-1506. DOI: [10.1007/s12010-014-1372-6](https://doi.org/10.1007/s12010-014-1372-6)
- Ryu, J., B. Nam, B.-R. Kim, S.H. Kim, Y.D. Jo, J.-W. Ahn, J.-B. Kim, C.H. Jin, and A.-R. Han , 2019. Comparative analysis of phytochemical composition of gamma-irradiated mutant cultivars of *Chrysanthemum morifolium*. *Molecules*, 24:3003. DOI: [10.3390/molecules24163003](https://doi.org/10.3390/molecules24163003)
- Saric, M., R. Kastrori, R. Curic, T. Cupina, and I. Gric, 1967. Chlorophyll Determination Univerzitet U Noveon Sadu Praktikum iz fiziologize Biljaka-Beograd. *Hauena Anjiga* 215.
- Shi, D.-Q., I. Ali, J. Tang, and W.-C. Yang, 2017. New insights into 5hmC DNA modification: generation, distribution and function. *Frontiers in genetics*, 8:100. <https://doi.org/10.3389/fgene.2017.00100>

- Silva, T.M., R.R. Miranda, V.P. Ferraz, M.T. Pereira, E.P. de Siqueira, and A.F. Alcântara, 2013. Changes in the essential oil composition of leaves of *Echinodorus macrophyllus* exposed to  $\gamma$ -radiation. *Revista Brasileira de Farmacognosia*, 23:600-607.  
DOI: [10.1590/S0102-695X2013005000049](https://doi.org/10.1590/S0102-695X2013005000049)
- Singh, B., and P.S. Datta, 2010. Effect of low dose gamma irradiation on plant and grain nutrition of wheat. *Radiation Physics and Chemistry*, 79:819-825.  
<https://doi.org/10.1016/j.radphyschem.2010.03.011>
- Studier, F.W., 1973. Analysis of bacteriophage T7 early RNAs and proteins on slab gels. *Journal of molecular biology*, 79:237-248. [https://doi.org/10.1016/0022-2836\(73\)90003-X](https://doi.org/10.1016/0022-2836(73)90003-X)
- Talská, R., J. Machalová, P. Smýkal, and K. Hron 2020. A comparison of seed germination coefficients using functional regression. *Applications in Plant Sciences*, 8:e11366.  
DOI: [10.1002/aps3.11366](https://doi.org/10.1002/aps3.11366)
- Tanabe, S., J. O'Brien, K.E. Tollefsen, Y. Kim, V. Chauhan, C. Yauk, E. Huliganga, R.A. Rudel, J.E. Kay, and J.S. Helm, 2022. Reactive Oxygen Species in the Adverse Outcome Pathway Framework: Toward Creation of Harmonized Consensus Key Events. *Frontiers in Toxicology* :81. <https://doi.org/10.3389/ftox.2022.887135>
- Telci, C., M. Yildiz, S. Pelit, B. Onol, E.G. Erkilic, and H. Kendir, 2011. The effect of surface-disinfection process on dormancy-breaking, seed germination, and seedling growth of *Lathyrus chrysanthus* Boiss. under in vitro conditions. *Propagation of Ornamental Plants*, 11:10-16.
- Thipyapong, P., M.D. Hunt, and J.C. Steffens, 1995. Systemic wound induction of potato (*Solanum tuberosum*) polyphenol oxidase. *Phytochemistry*, 40:673-676.  
[https://doi.org/10.1016/0031-9422\(95\)00359-F](https://doi.org/10.1016/0031-9422(95)00359-F)
- Tiwari, V., and D.M. Wilson III, 2019. DNA damage and associated DNA repair defects in disease and premature aging. *The American Journal of Human Genetics*, 105:237-257.  
DOI: [10.1016/j.ajhg.2019.06.005](https://doi.org/10.1016/j.ajhg.2019.06.005)
- Volkova, P.Y., G.T. Duarte, L. Soubigou-Taconnat, E.A. Kazakova, S. Pateyron, V.S. Bondarenko, S.V. Bitarishvili, E.S. Makarenko, R.S. Churyukin, and M.A. Lychenkova, 2020. Early response of barley embryos to low-and high-dose gamma irradiation of seeds triggers changes in the transcriptional profile and an increase in hydrogen peroxide content in seedlings. *Journal of Agronomy and Crop Science*, 206:277-295. <https://doi.org/10.1111/jac.12381>
- Wang, L., J. Wu, F. Lan, and P. Gao, 2020. Morphological, cytological and molecular variations induced by gamma rays in 'Donglinruixue'. *Folia Horticulturae*, 32:87-96.  
DOI: <https://doi.org/10.2478/fhort-2020-0009>
- Wang, P., Y. Zhang, L. Zhao, B. Mo, and T. Luo, 2017. Effect of gamma rays on *Sophora davidii* and detection of DNA polymorphism through ISSR marker. *BioMed research international*.  
DOI: [10.1155/2017/8576404](https://doi.org/10.1155/2017/8576404)
- Wi, S.G., B.Y. Chung, J.-S. Kim, J.-H. Kim, M.-H. Baek, J.-W. Lee, and Y.S. Kim, 2007. Effects of gamma irradiation on morphological changes and biological responses in plants. *Micron*, 38:553-564. DOI: [10.1016/j.micron.2006.11.002](https://doi.org/10.1016/j.micron.2006.11.002)
- Xiong, H., C. Zhou, H. Guo, Y. Xie, L. Zhao, J. Gu, S. Zhao, Y. Ding, and L. Liu, 2020. Transcriptome sequencing reveals hotspot mutation regions and dwarfing mechanisms in wheat mutants induced by  $\gamma$ -ray irradiation and EMS. *Journal of radiation research*, 61:44-57.  
<https://doi.org/10.1093/jrr/rrz075>
- Yue, Y., and J.M. Ruter 2020.  $^{60}\text{Co}$  irradiation influences germination and phenotype of three pavonia species. *HortScience*, 55:2037-2044. <https://doi.org/10.21273/HORTSCI15407-20>