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Changes in post-harvest life of cut chrysanthemum as influenced by different holding solutions and two cultivars

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ABSTRACT

To determine the appropriate treatment suitable for improving the heads flowers keeping quality and detecting some physiological changes occurring during the vase life period of the cut (Dendranthema grandiflorum, Ramat) cvs White Reagan and Arctic Queen heads flowers so, this study was consummated at post-Harvest. Lab. of floriculture Dep., Hort. Res. Inst., Giza, Egypt during 2013 and 2014 seasons to study the effect of some holding solutions viz. distilled water, (control), Gibberellic acid at 0.001%, Sucrose at (2 and 4 %), Citric acid at (150 mg/l), and 8-Hydroxygunoline citrate at (200 mg/l) on longevity of flower heads (vase life), water uptake, number of yellowing leaves and number of days from putting flowers in the different solutions to yellowing leaves (day). The obtained data exhibited that all holding solutions caused a marked increment in the studied characters compared to that registered from Sucrose (4%). In this respect, cut flowers treated with Sucrose at (2%) +8- Hydroxyqunoline citrate at (200 mg/l) + Gibberellic acid at (0.001 %) + Citric acid at (150 mg/l) decreased yellowing leaves and improved chlorophyll content and total carbohydrates in flower petals and leaves followed by the combined treatment of Sucrose at (4%) + 8-Hydroxyqunoline citrate at (200mg/l) + Gibberellic acid at (0.001%) + Citric acid at (150mg/l). Cut flowers (Dendranthema grandiflorum, Ramat.) cv. White Reagan gave the best results for studied characters compared to cv. Arctic Queen, results of interaction showed that all holding solutions had the highest effect on extending vase life, number of days taken for yellowing leaves and water uptake, while decreased number of yellowing leaves and improved chlorophyll content and total carbohydrates in flower petals and leaves .It could recommended to extend vase life of chrysantmemum cvs White Reagan and Arctic Queen holding cut flower in distilled water content sucrose (2%) + 8-Hydroxyqunoline citrate (8- HQC) (200 mg/l) + GA₃ (0.001 %)) + Citric acid (150 mg/l).

Keywords: Vase life, chrysanthemum cv. Whitr Reagan and Arctic Queen, Gibberellic acid, Sucrose, 8-Hydroxyquinoline citrate, Citric acid.

Introduction

The original home of Chrysanthemum is China, the species are of wide spread occurrence in temperate regions and in many parts of the globe, mostly in the old world. Chrysanthemum flowers are herbaceous perennial plants or subshrubs. They have alternately arranged leaves divided into leaflets with toothed or occasionally smooth edges. The compound inflorescence is an array of several flower heads, or sometimes a solitary head. The head has a base covered in layers of phyllaries. The simple row of ray florets is white, yellow, red. Cut flowers are parts of plants, characteristically including the blooms or "inflorescences" and some attached plant materials, but not including roots and soil. Fresh cut flowers are used for decorative purposes such as vase arrangements and boquets at formal event (Bonarriva, 2003).

Chrysanthemum (*Dendranthema grandiflora*.) is one of the most popular ornamental crops in the world (Saxena, 2001). It is a member of Asteraceae family; today, it is the second most economically important flower in the world after roses. (Nabigol *et al*, 2007). Cut flowers of chrysanthemum come in a wide range of colors and forms including two types namely, standard (one flower on the stem such as cv. "Arctic Queen" and "White Reagan" and spray (multiple flowers on the

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stem such as cv. "White Zambla"). The vase life is differing among various species and cultivars of chrysanthemum, which is one of the most valuable characteristics determining its quality, satisfying consumer preferences and the commercial value (Abou El-Ghait *et al*, 2012).

Vase life is an important parameter for evaluation of cut flower quality, for both domestic and export markets. The techniques of prolonging the vase life of flowers will be a great asset to the growers and users (Nair *et al*, 2003). The vase life is differing among various species and cultivars of chrysanthemum, which is one of the most valuable characteristics determining its quality, satisfying consumer preferences and the commercial value (Kumar, 2016).

The main postharvest problems in chrysanthemum is premature foliage yellowing, wilting and the failure of the flowers to fully open. The flowers of chrysanthemum have long vase life but its foliage often becomes yellow, brown and wilted during storage and transportation which reduces the postharvest quality of cut flowers. Foliage yellowing is cultivar specific and is caused by poor production, excessive or improper storage and preservative solutions used at higher than recommended concentrations. Leaves of cut chrysanthemum frequently become yellow spontaneously, sometimes prior to the onset of flower senescence, which makes the flowers unattractive, lowers their quality and shortens vase life (Doi *et al.*, 2003, 2004).

Adding chemical preservatives to the holding solution is recommended to preserve the best quality of flower after harvest. All holding solutions must contain sugars (Sucrose), acids (Citric acid) growth regulators, such Gibberellic acid and antimicrobials such as 8-Hydroxyquinoline (Abou El-Ghait *et al*, 2012).

Sugar play important role in plants as substances for respiration, the addition of sugars such as sucrose to vase water is effective in improving the vase life of cut flowers (Halevy and Mayak, 1979). Sucrose has been found to be the most commonly used sugar in prolonging vase life of cut flowers. The exogenous application of the sucrose supplies the cut flowers with much needed substrates for respiration, and enables cut flowers harvested at the bud stage to open, which otherwise could not occur naturally (Pun and Ichimura, 2003).

Also, Citric acid (CA) improved water balance and reduced stem plugging of bird of paradise cut flowers (Halevy *et al*, 1978), in addition applying Citric acid (CA) at (0, 100, 200 ppm) increased vase life, petal water content (%), initial fresh weight (%), marketability and significantly of chrysanthemum cut flowers (Vahdati Mashhadian *et al*, 2012).

Gibberellic acid prevented leaf chlorosis, which was the major postharvest disorder in many cut flowers such as Santonia cv. Golden light flowers (Eason *et al*, 2001).

8-HQS delayed senescence and eliminated bacterial growth which was the principal reason for reducing water uptake and transport of gerbera flower, (Abdel Kader, 1987).

This study aims to determine the appropriate treatment suitable for improving the flowers keeping quality and detecting some physiological changes occurring during the vase life period of cut (*Dendranthema grandiflorum*, Ramat) cvs. White Reagan and Arctic Queen heads flowers.

Material and Methods

The present study was conducted at the Ornamental Hort. Dep., Fac. Of Agric., Cairo Univ., Giza Egypt, during the period from 2012 to 2016.

This study had been carried out at Post-Harvest Lab. of Floriculture Dept., Hort. Res. Inst.; Agric. Res. Center, Ministry of Agric. and land reclamation, Giza, Egypt during 2013 and 2014 seasons. In order to determine the most suitable treatment for improving keeping quality and detecting some physiological changes taking place during the vase life period of cut chrysanthemum flowers.

Plant materials

Dendranthema grandiflorum (chrysanthemum) cvs. White Reagan and Arctic Queen cut flowers (standard: one flower on the stem). Cut flowers were obtained from a local commercial farm in Giza, Egypt in Jan. in both seasons. Chrysanthemum cut flowers were harvested in the early morning at their commercial stage which was full opening flowers Safa *et al.*, (2012) and wrapped in Kraft paper and transported to postharvest lab of Floriculture. Dept., Hort. Res. Inst., Giza, Egypt. During two hour. Cut flowers were precooled by placing in ice cold water for 2 hours to remove the effect of high

field heat. Stem bases were recut to about 3cm in air before treatments and adjusted to 60 cm long. 99 cut flowers of *Dendranthema grandiflorum* cvs. White Reagan and Arctic Queen were placed in a vase (500 ml) containing 300 ml holding solution to end vase life. All treatments were kept under condition i.e. 15-20°c, 60-70 % RH and continues light (1000-1500 lux) from white fluorescent.

Holding solutions:

- 1- Distilled water (D.W), as control = T_1 .
- 2- Sucrose (Suc.) $(2\%) = T_2$.
- 3- Sucrose $(4\%) = T_3$.
- 4- Sucrose (2%) + 8-Hydroxyqunoline citrate (8-HQC) (200 mg/l) = T_4 .
- 5- Sucrose (4%) + 8- Hydroxyqunoline citrate (200 mg/l)= T_5 .
- 6- Sucrose (2%) + Gibberellic acid (GA₃) (0.001 %) = T_6 .
- 7- Sucrose (4%) + Gibberellic acid (0.001 %) = T_7 .
- 8- Sucrose (2%) + 8- Hydroxyqunoline citrate (200 mg/l) + Gibberellic acid (0.001%) + Citric acid (150 mg/l)= T_8 .
- 9- Sucrose (2%) + Citric acid (150 mg/l)= T_9 .
- 10- Sucrose (4%) + Citric acid (150 mg/l)= T_{10} .
- 11- Sucrose (4%) +8- Hydroxyqunoline citrate (200 mg/l) + Gibberellic Acid (0.001%) + Citric Acid (150 mg/l)= T₁₁.

1. Data record

- Vase life (day): day number till wilting.
- Water uptake (ml/3 flowers): (the solution volume at the beginning of the study- the solution volume of 1st, 8th, 15th, 22th, 30th day in *Dendranthema grandiflorum cvs*. White Reagan and Arctic Queen cut flowers (ml/3 flowers).
- Number of yellowing leaves.
- Number of days from putting flowers in different solutions to yellowing leaves (day).

2. Chemical analysis:

- Total carbohydrates (%) in the flower petals were determined calorimetrically, according to the methods described by Dubois *et al*, (1956).
- Chlorophyll a,b and carotenoids content (mg/g) in flower petals was determined colorimetrically at the end of storage period according to Husia *et al*, (1965).

3. Layout of the experiment

The experimental layout was a complete randomized design in a factorial experiment (two factors). Holding solutions (factor A) was applied in 11 solutions and two cultivars on (factor B).

Statistical analysis

The data were statistically analyzed as a factorial experiment using MSTAT-C (1993) and means were compared by Duncan's Multiple Rang Test as described by Duncan (1955) to verify differences among means of various treatments.

Results and Discussion

Effect of different cultivars, holding solutions and their interactions on quality of (*Dendranthema grandiflorum*, Ramat.) cvs. White Reagan and Arctic Queen cut flowers during 2013 and 2014:

1. Flower vase life:

Data presented in Table (1) reveal that the significantly highest vase life was obtained with *Dendranthema grandiflorum* cv. White Reagan (22.36 and 25.63days) compared to the second cultivars Arctic Queen (19.36 and 20.88 days), in first and second seasons respectively.

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Table 1: Effect of different cultivars on vase life (days) of (*Dendranthema grandiflorum*, Ramat) cvs White Reagan and Arctic Queen cut flowers during 2013 and 2014.

Cultivars	1st season 2013	2 nd season 2014
White Reagan	22.36a	25.63a
Arctic Queen	19.36b	20.88b

Means within column having the same letters are not significantly different according to Ducan's multiple range test

Regarding the effect of holding solutions treatments on vase life, data presented in Table (2) indicate that all treatments increased vase life of *Dendranthema grandiflorum*m cvs. White Reagan and Arctic Queen in the two seasons compared to (T_1) distilled water , $(T_2$ and $T_3)$ Sucrose (2 and 4%). Data show that treating flowers with (T_8) Suc (2%) + 8-HQC $(200 \text{ mg/l}) + GA_3 (0.001\%) + CA (150 \text{ mg/l})$ gave the highest significantly effect on longevity (31.07 and 33.43) followed by the last solution containing with (T_{11}) Suc (4%) + 8-HQC $(200 \text{ mg/l}) + GA_3 (0.001\%) + CA (150 \text{mg/l}) (28.75 \text{ and } 31.60)$ in the both seasons respectively.

Table 2: Effect of holding solutions on vase life (days) of (*Dendranthema grandiflorum*, Ramat.) evs White Reagan and Arctic Queen cut flowers during 2013 and 2014.

Holding solutions treatments	1 st season	2 nd season
	2013	2014
T ₁	15.23i	17.22i
T_2	12.33j	15.05j
T_3	9.983k	13.07k
T ₄	26.72c	29.32c
T ₅	25.45d	27.10d
T_6	19.28g	21.43g
T_7	17.38h	19.52h
T_8	31.07a	33.43a
T 9	23.38e	25.03e
T_{10}	20.87f	23.05e
T_{11}	28.75b	31.60b

Means within column having the same letters are not significantly different according to Ducan's multiple range test

 T_1 = Distilled water T_2 = Sucrose (2%)

T3= Sucrose (4%)

T4 = Sucrose (2%) + 8-Hydroxyqunoline citrate (200mg/l)

 $\textbf{T5} = \text{Sucrose } (4\%) + 8 + \text{Hydroxyqunoline citrate } (200 \text{ mg/l}) \\ \textbf{T6} = \text{Sucrose } (2\%) + \text{Gibbrillic acid } (0.001\%) \\ \textbf{T6} = \text{Sucrose } (2\%) + \text{Gibbrillic acid } (0.001\%) \\ \textbf{T6} = \text{Sucrose } (2\%) + \text{Gibbrillic acid } (0.001\%) \\ \textbf{T6} = \text{Sucrose } (2\%) + \text{Gibbrillic acid } (0.001\%) \\ \textbf{T6} = \text{Sucrose } (2\%) + \text{Gibbrillic acid } (0.001\%) \\ \textbf{T6} = \text{Sucrose } (2\%) + \text{Gibbrillic acid } (0.001\%) \\ \textbf{T6} = \text{Sucrose } (2\%) + \text{Gibbrillic acid } (0.001\%) \\ \textbf{T6} = \text{Sucrose } (2\%) + \text{Gibbrillic acid } (0.001\%) \\ \textbf{T6} = \text{Sucrose } (2\%) + \text{Gibbrillic acid } (0.001\%) \\ \textbf{T6} = \text{Sucrose } (2\%) + \text{Gibbrillic acid } (0.001\%) \\ \textbf{T6} = \text{Sucrose } (2\%) + \text{Gibbrillic acid } (0.001\%) \\ \textbf{T6} = \text{Sucrose } (2\%) + \text{Gibbrillic acid } (0.001\%) \\ \textbf{T6} = \text{Sucrose } (2\%) + \text{Gibbrillic acid } (0.001\%) \\ \textbf{T6} = \text{Sucrose } (2\%) + \text{Gibbrillic acid } (0.001\%) \\ \textbf{T7} = \text{Gibbrillic acid } (0.001\%) \\ \textbf{T8} = \text{Gibbrillic acid$

T7 = Sucrose (4%) + Gibbrillic acid (0.001%)

T8= Sucrose (2%) + 8-Hydroxyqunoline citrate(200mg/l) + Gibbrillic acid (0.001%) + Citric acid (150mg/l)

T9 = Sucrose (2%) + Citric acid (150mg/l) T10 = Sucrose (4%) + Citric acid (150mg/l)

T11= Sucrose (4%) + 8-Hydroxyqunoline citrate (200mg/l) + Gibbrillic acid (0.001%) + Citric acid (150mg/l)

Data in Table (3) show that treatment with (T_8) Suc (2 %) + 8-HQC $(200 \text{ mg/l}) + \text{GA}_3(0.001 \%) + \text{CA}$ (150 mg/l) was the most effective treatment for prolonging longevity with the two cultivars followed by cut flowers treated with (T_{11}) (Suc 4%) + 8-HQC $(200 \text{ mg/l}) + \text{GA}_3 (0.001\%) + \text{CA} (150 \text{ mg/l})$ compared with other treatments in the first and second seasons respectively.

These results are in accordance with those reported by Sharma *et al*, (2005) who found that the highest vase life of *lilium grandiflorum* cut flowers was obtained with (2%) Sucrose + (50 ppm) GA₃+ (200 ppm) 8-HQC, also Li *et al*, (2012) mentioned that the vase life of cut carnations increased when holding (5%) Sucrose + (100 mg/l) 8-HQ + (200 mg/l) CA + (100mg/l) 6-BA. Abd El-Kafie *et al*, (2014) investigate that the chrysanthemum cut flowers were held in the vase solution containing Silver nitrate (75 ppm) + Citric acid (150 ppm) + Sucrose at (3%) was increased significant vase life (19.5 and 17.2 days, respectively). Preservative solutions containing (400 ppm) 8-HQC and (1.5%) Sucrose gave that maximum vase life of chrysanthemum (*Dendranthema grandiflora*, Ramat) cv. White Reagan cut flowers (Jain *et al*, 2014).

2. Number yellowing leaves (%):

Data in Table (4) clear that the number of yellowing leaves increased in cut flowers *Dendranthema grandiflorum* cv. Arctic Queen (3.04 and 2.82%) when compared with cv. White Reagan (2.69 and 2.33%) in both seasons respectively.

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Table 3: Effect of interaction between different cultivars, holding solutions on vase life of (Dendranthema grandiflorum, Ramat.) cvs. White Reagan and Arctic Queen cut flowers during 2013 and 2014.

441115 2010 WH 2011.	Cul	tivars of <i>Dendra</i>	nthema grandiflo	rum
Holding solutions treatments	20	13	20	14
	White	Arctic	White	Arctic
	Reagan	queen	Reagan	queen
T ₁ T ₂	16.571	13.90m	19.231	15.20n
	13.00m	11.67n	17.10m	13.00o
T ₃	10.200	9.760	15.23n	10.90p
T ₄ T ₅	28.00cd	25.43f	32.00c	26.63g
	27.00de	23.90g	29.43e	24.77hi
T ₆	20.70i	17.87k	23.77i	19.101
T ₇	19.00j	15.771	21.13jk	17.90m
	33.77a	28.37c	36.10a	30.77d
T ₈ T ₉	24.87fg	28.37C 21.90h	27.87f	22.20j
T_{10} T_{11}	22.10h	19.63ij	25.67gh	20.43k
	30.80b	26.70e	34.43b	28.77ef

Means within column having the same letters are not significantly different according to Ducan's multiple range test

T₁= Distilled water

T2 = Sucrose (2%)

T₃= Sucrose (4%) T5 = Sucrose (4%) + 8-Hydroxyqunoline citrate (200 mg/l)

T7 = Sucrose (4%) + Gibbrillic acid (0.001%)

T9 = Sucrose (2%) + Citric acid (150mg/l)

T4 = Sucrose (2%) + 8-Hydroxyqunoline citrate (200mg/l)

T6 = Sucrose (2%) + Gibbrillic acid (0.001%)

T8= Sucrose (2%) + 8-Hydroxyqunoline citrate(200mg/l) +

Gibbrillic acid(0.001%) + Citric acid (150mg/l)

T10 = Sucrose (4%) + Citric acid (150mg/l)T11= Sucrose (4%) + 8-Hydroxyqunoline citrate (200mg/l) + Gibbrillic acid (0.001%) + Citric acid (150mg/l)

Table 4: Effect of the different cultivars on number of yellowing leaves (%) of (Dendranthema grandiflorum, Ramat.) cvs. White Reagan and Arctic Oueen cut flowers during seasons 2013 and 2014.

/	C		C
Cultivars		1 st season 2013	2 nd season 2014
White Reagan		2.69b	2.33b
Arctic Queen		3.04a	2.82a

Means within column having the same letters are not significantly different according to Ducan's multiple range test.

Data in Table (5) indicate that all holding solutions significantly decreased the number of yellowing leaves of dendranthema cut flowers meanwhile treating cut flowers with (T₁) distilled water, (T₂) sucrose 2%, (T₃) sucrose 4% and (T₁₀) Suc 4% + CA 150mg/l increased number of yellowing leaves in both seasons.

Concerning the data of interaction (holding solutions x different cultivars) the obtained data in Table (6) mentioned that treating cut flowers in 2 cultivars with different holding solutions decreased the number of yellowing leaves, compared with (T₁) distilled water, (T₃ and T₂) Sucrose (4% and 2%) in the two seasons.

This agreed with the results found by Han (2001) who stated that the yellowing leaves decreased in cut lilies when used high concentrations of growth regulators like GA₃. Using Gibberellic acid treatment (20, 50, and 100 mg/l) in combination with Sucrose (4%) and Citric acid (200 mg/l) prevented the yellowing of lily leaves of the cut flowers of Lilium longiflorum. (Dias-Tagliacozzo et al (2005), Also Jain et al (2014) showed that reduce the foliage discoloration of chrysanthemum (Dendranthema grandiflora, Ramat.) cv. White Reagan when held flowers in preservative solutions containing (400ppm) 8-HQC and (1.5%) sucrose.

3- Number of days from putting flowers in different solutions to yellowing leaves (day).

Data presented in Table (7) evident that the *Dendranthema grandiflorum* cv. White Reagan cut flowers gave highest number of days for yellowing leaves (15.94 and 19.11days) than cv. Arctic Queen cut flowers (13.38 and 15.28 days), in both seasons respectively.

Table 5: Effect of holding solutions on number of yellowing leaves (%)on (*Dendranthema grandiflorum*, Ramat.) cvs. White Reagan and Arctic Queen during seasons 2013 and 2014.

H-11'	1st season	2 nd season
Holding solutions treatments	2013	2014
T ₁	4.21c	3.81c
T ₂	4.93b	4.60b
T ₃	5.68a	5.33a
T ₄	2.58e	2.21e
T ₅	2.83e	2.50e
T_6	1.23fg	1.00f
T ₇	1.45f	1.21f
T ₈	0.50h	0.35g
T ₉	3.56d	3.16d
T ₁₀	8.78cd	3.50cd
T ₁₁	0.78gh	0.66fg

Means within column having the same letters are not significantly different according to Ducan's multiple range test

T₁= Distilled water

T2 = Sucrose (2%)

T3= Sucrose (4%)

T4 = Sucrose (2%) + 8-Hydroxyqunoline citrate (200mg/l)

T5 = Sucrose (4%) + 8-Hydroxyqunoline citrate (200 mg/l)

T6 = Sucrose (2%) + Gibbrillic acid (0.001%)

T7 = Sucrose (4%) + Gibbrillic acid (0.001%)

T8= Sucrose (2%) + 8-Hydroxyqunoline citrate(200mg/l) +

Gibbrillic acid(0.001%) + Citric acid (150mg/l)

T9 = Sucrose (2%) + Citric acid (150mg/l)

T10 = Sucrose (4%) + Citric acid (150mg/l)

T11= Sucrose (4%) + 8-Hydroxyqunoline citrate (200mg/l) + Gibbrillic acid (0.001%) + Citric acid (150mg/l)

Table 6: Effect of interaction between different cultivars, holding solutions on number of yellowing leaves (%) on (*Dendranthema grandiflorum*, Ramat.) cvs. White Reagan and Arctic Queen during seasons 2013 and 2014.

	Cultivars of Dendranthema grandiflorum							
Holding solutions treatments	20	13	20	14				
	White Reagan	Arctic queen	White Reagan	Arctic queen				
T1	3.90cd	4.53bc	3.43cde	4.20bc				
T2	4.76b	5.10ab	4.33bc	4.86ab				
Т3	5.56a	5.80a	5.00ab	5.66a				
T4	2.36f	2.80ef	2.00gh	2.43fg				
T5	2.53f	3.13def	2.20gh	2.80efg				
T6	1.00ghi	1.47gh	0.90ij	1.10ij				
T7	1.33gh	1.57g	1.00ij	1.43hi				
T8	0.33i	0.67hi	0.23j	0.47j				
Т9	3.46de	3.67d	2.80efg	3.53cde				
T10	3.66d	3.90cd	3.23ef	3.77cd				
T11	0.67hi	0.90ghi	0.53ij	0.80ij				

Means within column having the same letters are not significantly different according to Ducan's multiple range test

T₁= Distilled water

T2 = Sucrose (2%)

T3= Sucrose (4%)

T4 = Sucrose (2%) + 8-Hydroxyqunoline citrate (200mg/l)

T5 = Sucrose (4%) + 8-Hydroxyqunoline citrate (200 mg/l)

T6 = Sucrose (2%) + Gibbrillic acid (0.001%)

T7 = Sucrose (4%) + Gibbrillic acid (0.001%)

T9 = Sucrose (2%) + Citric acid (150mg/l)

T8= Sucrose (2%) + 8-Hydroxyqunoline citrate(200mg/l) +

Gibbrillic acid(0.001%) + Citric acid (150mg/l)

T10 = Sucrose (4%) + Citric acid (150 mg/l)

T11= Sucrose (4%)+8-Hydroxyqunoline citrate (200mg/l) + Gibbrillic acid (0.001%) + Citric acid (150mg/l)

Table7: Effect of the different cultivars on number of days from putting flowers in different solutions to yellowing leaves of *(Dendranthema grandiflorum, Ramat.)* cvs. White Reagan and Arctic Queen cut flowers during seasons 2013 and 2014.

Cultivars	1st season 2013	2 nd season 2014
White Reagan	15.94a	19.11a
Arctic Queen	13.38b	15.28b

Means within column having the same letters are not significantly different according to Ducan's multiple range test

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Data in Table (8) show that treating dendranthema cut flowers with different holding solutions increased the number of days for yellowing leaves, while treating them with (T_8) Suc (2 %) + 8-HQC $(200 \text{ mg/l}) + \text{GA}_3 (0.001 \%) + \text{CA} (150 \text{ mg/l})$ was increased the number of days for yellowing leaves, followed by treating with (T_{11}) Sucrose (4 %) + 8-HQC (200 mg/l) + GA3 (0.001 %) + CA (150 mg/l) compared with the other treatments in both seasons.

Table 8: Effect of holding solutions on number of days from putting flowers in different solutions to yellowing leaves of (*Dendranthema grandiflorum*, Ramat.) cvs. White Reagan and Arctic Queen cut flowers during seasons 2013 and 2014.

Holding solutions treatments	1 st season	2 nd season	
	2013	2014	
T1	8.73i	11.18i	
T2	6.67j	9.27j	
T3	5.67k	7.45k	
T4	13.95e	17.17e	
T5	13.02f	15.77f	
T6	17.95c	20.72c	
T7	15.62d	18.00d	
T8	30.38a	33.00a	
T9	11.38g	13.78a	
T10	9.93h	12.38h	
T11	27.95b	30.43b	

Means within column having the same letters are not significantly different according to Ducan's multiple range test

T₁= Distilled water

T3= Sucrose (4%)

T5 = Sucrose (4%) + 8-Hydroxyqunoline citrate (200 mg/l)

T7 = Sucrose (4%) + Gibbrillic acid (0.001%)

T9 = Sucrose (2%) + Citric acid (150mg/l)

T4 = Sucrose (2%) + 8-Hydroxyqunoline citrate (200mg/l) T6 = Sucrose (2%) + Gibbrillic acid (0.001%)

T8= Sucrose (2%) + 8-Hydroxyqunoline citrate(200mg/l) +

Gibbrillic acid(0.001%) + Citric acid (150mg/l) T10 = Sucrose (4%) + Citric acid (150mg/l)

 $T_2 = Sucrose (2\%)$

T11= Sucrose (4%) + 8-Hydroxyqunoline citrate (200mg/l) + Gibbrillic acid (0.001%) + Citric acid (150mg/l)

The effect of the interaction between holding solutions treatments and different cultivars from dendranthema cut flowers is shown in Table (9). Data indicate that the highest number of days for yellowing leaves was obtained by supplemented with (T_8) Suc (2%) + 8-HQC (200 mg/l) + GA_3 (0.001%) + GA_3 (0.001%) + GA_3 (0.001%) + GA_3 (0.001%) + GA_3 (150 mg/l) in cv. White Reagan compared with other treatments, in the first and second seasons. The present results are also in accordance with those reported by Jain *et al*, (2014) who mentioned that the maximum number of days taken for yellowing leaves of *Chrysanthemum grandiflora* cv. White Reagan cut flower was recorded in the treatment containing 8-HQC (400ppm) and sucrose (1.5%) as the holding solution.

4- Water uptake:

The effect of the difference between dendranthema cultivars is shown in Table (10) it is clear that *Dendranthema grandiflorum* cv. White Reagan raised the amount of water uptake till the 22th day (492.58, 601.52) and also, cv. Arctic Queen raised the amount of water uptake till the 22th day (425.45, 478.33) in the first and second season respectively.

Data presented in Table (11) clearly indicate that holding solutions containing of (T_8) Suc (2%) + 8-HQC (200 mg/l) + GA₃ (0.001 %) + CA (150 mg/l) was the best treatment for increasing water uptake of dendranthema cut flowers till the 30th day in the first and the second season. Meanwhile dendranthema cut flowers with (T_1) distilled water increased water uptake till the 15th day in the both season while (T_3 and T_2) sucrose (4 and 2%) was increased water uptake till the 8th day in the first season and 15th day in the second one.

According to data presented in Table (12) it is obvious that treating *Dendranthema grandiflorum* cv. White Reagan cut flowers with (T_8) Suc at (2 %) + 8-HQC (200 mg/l) + GA₃ (0.001 %) + CA (150 mg/l) was increased water uptake till the 30th day in the first season and the second season, compared with other treatments. This matches with Kumar (2016) who mentioned that treating chrysanthemum cut flowers with different solutions content of sucrose, citric acid and 8-HQC increasing the flowers water uptake, also Zamani *et al* (2011) showed that sucrose at 3 %, citric acid, malic acid and salicylic acid increasing water uptake in chrysanthemum cut flowers.

Table 9: Effect of interaction between different cultivars and holding solutions on number of days from putting flowers in different solutions to yellowing leaves of (Dendranthema grandiflorum, Ramat.) cvs. White Reagan and Arctic Queen cut flowers during Seasons 2013 and 2014.

	Cultivars of Dendranthema grandiflorum						
Holding solutions treatments	20	13	2	014			
Holding solutions treatments	White	Arctic	White	Arctic			
	Reagan	queen	Reagan	queen			
T1	9.80jk	7.671	12.80k	9.57mn			
T2	7.431	5.90m	10.871	7.67o			
Т3	6.90lm	4.43n	8.90n	6.00p			
T4	15.00g	12.90h	19.00fg	15.33ij			
T5	14.23g	11.80hi	17.20h	14.33j			
T6	18.90e	17.00f	23.20e	18.23gh			
T7	16.33f	14.90g	20.00f	16.00i			
T8	33.10a	27.67c	35.77a	30.23c			
T9	12.77h	10.00jk	15.00ij	12.57k			
T10	10.90ij	8.97k	14.23j	10.53lm			
T11	30.00b	25.90d	33.20b	27.67d			

Means within column having the same letters are not significantly different according to Ducan's multiple range test

T₁= Distilled water

T3= Sucrose (4%)

T5 = Sucrose (4%) + 8-Hydroxyqunoline citrate (200 mg/l)

T7 = Sucrose (4%) + Gibbrillic acid (0.001%)

T9 = Sucrose (2%) + Citric acid (150mg/l)

 $T_2 = Sucrose (2\%)$

T4 = Sucrose (2%) + 8-Hydroxyqunoline citrate (200mg/l)

T6 = Sucrose (2%) + Gibbrillic acid (0.001%)

T8= Sucrose (2%) + 8-Hydroxyqunoline citrate(200mg/l) +

Gibbrillic acid(0.001%) + Citric acid (150mg/l) T10 = Sucrose (4%) + Citric acid (150 mg/l)

T11= Sucrose (4%) + 8-Hydroxyqunoline citrate (200mg/l) + Gibbrillic acid (0.001%) + Citric acid (150mg/l)

Table 10: Effect of the different cultivars on water uptake (ml/3 flowers) of (Dendranthema grandiflorum, Ramat.) cvs. White Reagan and Arctic Queen cut flowers during seasons 2013 and 2014.

		1st season 2013	3		
Cultivars	1 st	8 th	15 th	22 th	30 th
White Reagan	38.99a	267.73a	449.09a	492.58a	356.67a
Arctic queen	36.364b	219.09b	404.55b	425.45b	252.58b
-		2 nd season 201	4		
Cultivars	1 st	8 th	15 th	22 th	30 th
White Reagan	48.333a	287.88a	508.33a	601.52a	484.24a
Arctic queen	42.121b	234.39b	441.36b	478.33b	263.33b

Means within column having the same letters are not significantly different according to Ducan's multiple range test

Table 11. Effect of holding solutions on water uptake (ml / 3flowers) of chrysanthemum (Dendranthema grandiflorum, Ramat.) cvs. White Reagan and Arctic Queen cut flowers during seasons 2013 and 2014.

Holding solutions treatments	1st season 2013						2 nd season 2014			
treatments	1 st	8 th	15 th	22 th	30^{th}	1 st	8 th	15 th	22 th	30^{th}
T1	31.94d	188.33h	343.33e			41.66d	195.00g	353.33f	258.33g	
T2	28.33e	166.67i	103.33f			38.33e	182.50g	277.50g		
Т3	25.83e	145.00j				36.67e	160.0h	157.50h		
T4	44.17a	295.00c	556.67b	769.17a-c	807.50a	50.83b	315.83c	592.50b	789.17a-c	875.00a
T5	40.00b	276.67d	525.83bc	723.33b-d	520.00b	46.67c	295.83d	544.17c	758.33b-d	575.0b
T6	35.83c	222.50f	441.67d	414.17e		42.50d	231.67f	476.67de	628.33e	128.33de
T7	35.00c	202.50g	428.33d	154.17f		42.50d	222.50f	442.50e	409.17f	
T8	46.66a	363.33a	645.83a	841.67a	924.17a	54.17a	358.00a	684.17a	863.33a	970.83a
Т9	40.83b	252.50e	518.33bc	686.67cd	224.17c	45.83c	282.50d	531.67c	711.67cd	406.67bc
T10	40.00b	232.50f	503.33c	651.67d		45.83c	258.33e	518.33cd	686.67de	255.00cd
T11	45.83a	327.50b	628.33a	808.33a-b	875.00a	52.50ab	343.3b	645.00a	834.17ab	900.83a

Means within column having the same letters are not significantly different according to Ducan's multiple range test

T1= Distilled water **T3**= Sucrose (4%)

T2 = Sucrose (2%)

T5 = Sucrose (4%) + 8-Hydroxyqunoline citrate (200 mg/l)

T4 = Sucrose (2%) + 8-Hydroxyqunoline citrate (200mg/l) T6 = Sucrose (2%) + Gibbrillic acid (0.001%)

T7 = Sucrose (4%) + Gibbrillic acid (0.001%)

T8= Sucrose (2%) + 8-Hydroxygunoline citrate(200mg/l) +

Gibbrillic acid(0.001%) + Citric acid (150mg/l)

T10 = Sucrose (4%) + Citric acid (150mg/l)

T9 = Sucrose (2%) + Citric acid (150mg/l)T11= Sucrose (4%) + 8-Hydroxyqunoline citrate (200mg/l) + Gibbrillic acid (0.001%) + Citric acid (150mg/l)

Table 12: Effect of interaction between different cultivars, holding solutions on water uptake (ml / 3 flowers) of (*Dendranthema grandiflorum*, Ramat.) cvs. White Reagan and Arctic Queen cut flowers during Seasons 2013 and 2014.

	Cultivars of <i>Dendranthema grandiflorum</i> 1 st season 2013									
Holding solutions		1 st	8	th		5 th	22 th		30 th	
treatments	White reagan	Arctic queen	White reagan	Arctic queen	White reagan	Arctic queen	White reagan	Arctic queen	White reagan	Arctic queen
T1	33.89f	30.00g	223.33ij	153.33m	366.67ij	320.00j				
T2	30.00g	26.67g	205.00k	128.33n	206.67k					
T3	30.00g	21.67h	180.001	110.00o						
T4	45.00bc	43.33b-d	316.67c	273.33e	573.33bc	540.00cd	785.00a-c	753.33a-d	823.33a	791.67a
T5	40.00de	40.00de	296.67d	256.67fg	541.67cd	510.00с-е	743.33a-d	703.33b-d	783.33a	256.67b
T6	36.67ef	35.00f	246.67gh	198.33k	456.67e-h	426.67g-i	518.33e	310.00f		
T7	35.00f	35.00f	231.67hi	173.331	440.00f-h	416.67hi	308.33f			
T8	50.00a	43.33b-d	383.33a	353.33b	661.67a	630.00ab	853.33a	830.00ab	948.33a	900.00a
Т9	41.67cd	40.00de	270.00ef	235.00hi	533.33cd	503.33d-f	706.67b-d	666.67cd	448.33b	
T10	40.00de	40.00de	253.33g	211.67jk	516.67с-е	490.00d-g	683.33cd	620.00de		
T11	43.37ab	45.00bc	338.33b	316.67c	643.33a	613.33ab	820.00ab	796.67a-c	920.00a	830.00a
					2 nd season 2	014				
T1	43.33с-е	40.00ef	228.33hi	161.671	375.00kl	331.671	516.67i			
T2	40.00ef	36.67fg	221.67h-j	143.33lm	348.331	206.67m				
Т3	40.00ef	33.33g	195.00k	125.00m	315.001					
T4	55.00a	46.67bc	348.33b	283.33de	606.67b-d	578.33с-е	818.33a-d	760.00b-f	913.33a	836.67a
T5	50.00b	43.33с-е	321.67c	270.00ef	565.00d-f	523.33e-h	783.33a-e	733.33с-д	890.00a	260.00bc
T6	45.00cd	40.00ef	251.67fg	211.67i-k	495.00g-j	458.33h-j	666.67f-h	590.00hi	256.67bc	
T7	45.00cd	40.00ef	243.33gh	201.67jk	448.33ij	436.67jk	635.00gh	183.33j		
T8	58.33a	50.00b	406.67a	363.33b	705.00a	663.33ab	886.67a	840.00a-c	995.00a	946.67a
Т9	50.00b	41.67de	296.67d	268.33ef	550.00d-g	513.33e-i	733.33с-д	690.00e-H	813.33a	
T10	50.00b	41.67de	286.67de	230.00g-i	528.33e-g	508.33f-i	710.00d-G	663.33f-H	510.00b	
T11	55.00a	50.00b	366.67b	320.00c	655.00ab	635.00bc	866.67ab	801.67a-e	948.33a	853.33a

Means within column having the same letters are not significantly different according to Ducan's multiple range test

T₁= Distilled water

T3= Sucrose (4%)

T5 = Sucrose (4%) + 8-Hydroxyqunoline citrate (200 mg/l)

T7 = Sucrose (4%) + Gibbrillic acid (0.001%)

T9 = Sucrose (2%) + Citric acid (150mg/l)

T2 = Sucrose (2%)

T4 = Sucrose (2%) + 8-Hydroxyqunoline citrate (200mg/l)

T6 = Sucrose (2%) + Gibbrillic acid (0.001%)

T8= Sucrose (2%) + 8-Hydroxyqunoline citrate(200mg/l) + Gibbrillic acid(0.001%) + Citric acid (150mg/l)

T10 = Sucrose (4%) + Citric acid (150mg/l)

T11= Sucrose (4%) + 8-Hydroxyqunoline citrate (200mg/l) + Gibbrillic acid (0.001%) + Citric acid (150mg/l)

Total carbohydrates %:

Data presented in Fig.(1) illustrated that the carbohydrates % increased in *Dendranthema grandiflorum* cv White Reagan, compared to cv Arctic Queen (in leaves and petals) in both seasons.

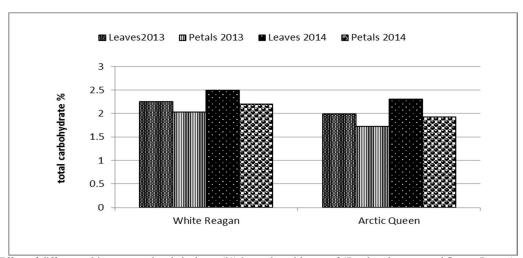


Fig. 1: Effect of different cultivars on total carbohydrates (%) in petals and leaves of (*Dendranthema grandiflorum*, Ramat.) cvs. White Reagan and Arctic Queen during seasons 2013 and 2014

Regarding the effect of holding solutions, it was concluded from data in Fig. (2) That, the dendranthema cut flowers treated with (T_3 and T_2) Suc (4 and 2%) and(T_1) distilled water had the least values of the carbohydrates % compared to all the other treatments used in the first and second

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seasons. The highest values of the carbohydrates % were found in the dendranthema cut flowers treated with (T_8) Suc (2%) + 8-HQC (200mg/l) + GA₃ (0.001%) + CA (150 mg/l) followed by cut flowers treated with (T_{11}) Suc at (4%) + 8-HQC (200mg/l) + GA₃ (0.001%) + CA (150mg/l) as compared to other treatments used in the first and second seasons.

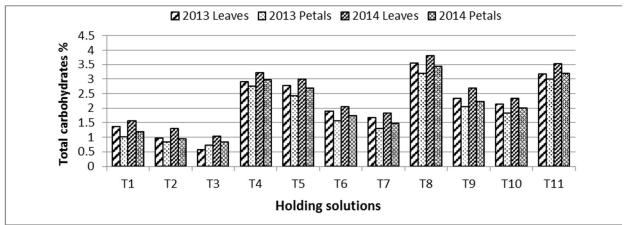


Fig. 2: Effect of holding solutions on total carbohydrate (%) in petals and leaves of (Dendranthema grandiflorum, Ramat) cvs. White Reagan and Arctic Queen during seasons 2013 and 2014

T₁= Distilled water

T3= Sucrose (4%)

T5 = Sucrose (4%) + 8-Hydroxyqunoline citrate (200 mg/l)

T7 = Sucrose (4%) + Gibbrillic acid (0.001%)

T4 = Sucrose (2%) + 8-Hydroxyqunoline citrate (200mg/l) T6 = Sucrose (2%) + Gibbrillic acid (0.001%)

T2 = Sucrose (2%)

T8= Sucrose (2%) + 8-Hydroxyqunoline citrate(200mg/l) + Gibbrillic acid(0.001%) + Citric acid (150mg/l)

T10 = Sucrose (4%) + Citric acid (150mg/l)T9 = Sucrose (2%) + Citric acid (150mg/l)T11= Sucrose (4%) + 8-Hydroxyqunoline citrate (200mg/l) + Gibbrillic acid (0.001%) + Citric acid (150mg/l)

The interaction between holding solutions treatments and different cultivars from dendranthema cut flowers as shown in fig (3) and fig (4), which indicate that Dendranthema grandiflorum cv White Reagan held in solution containing (T₈) Suc (2%) + 8-HQC (200mg/l) + GA₃ (0.001%) + CA (150mg/l) were the best for obtaining the highest carbohydrates % as compared to the other treatments. The previous results are in agreement with Hwang et al, (2009) on Lilium oriental hybride "Casa Blanca" he found that the holding solution containing (3%) Sucrose + (200mg/l) HQC +(25 mg/l) GA₃ maintained high levels of total sugar for a long time as compared with the control.

Amin (2017) mentioned that the maximum content in total soluble sugar for chrysanthemum cut flowers cv. Arctic Queen when treated with combination of (Sucrose + Citric acid), also Su et al, (1991) reported in chrysanthemum cut flowers treated with (5%) Sucrose + (150ppm) Citric acid + (50 ppm) AgNo₃ increased the soluble sugar content in the petal.

Chlorophyll content (mg/g):

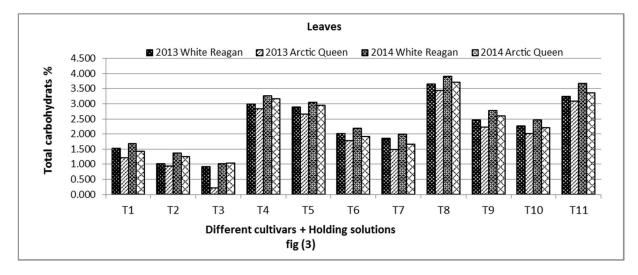
Data presented in Fig. (5) Indicate that Dendranthema grandiflorum cv White Reagan gave higher values of chlorophyll content than Chrysanthemum grandiflorum cv Arctic queen.

Data illustrated in Fig.(6) show that the treatment with (T₃ and T₂) Suc (4, 2%) and (T₁) distilled water gave the least values of chlorophyll content (mg/g) in the leaves of dendranthema cut flowers compared to other treatments in both seasons. However, treated flowers with (T₈) Suc (2%) + 8-HQC (200mg/l) + GA₃ (0.001%) + CA (150 mg/l) recorded the highest values of chlorophyll content (mg/g) followed by treating with (T_{11}) Suc (4%) + 8-HQC (200mg/l) + GA_3 (0.001%) + CA (150mg/l) in both seasons.

Data in Fig. (7,8 and 9) showed that the Dendranthema grandiflorum cv White Reagan cut floweres treated with (T_8) Suc (2%) + 8-HQC (200 mg/l) + GA3 (0.001%) + CA (150 mg/l) gaves the highest value of chlorophyll content (mg/g) in leaves compared with all other treatments, in the two seasons.

These results are in accordance with those of Gendy (2000) who indicate that Gladiolus cut flowers treated with holding solutions containing Sucrose (10%) + 8-HQ (200ppm)+ Citric acid (150ppm) was most effective for increasing chlorophyll content when compared to the other treatment. Emami et al. (2011) mentioned that the highest chlorophyll a content was related to

Gibberllic acid at (75 ppm) in (*Lilium longiflorum*) cut flowers. Also, Osman and Sewedan (2014) showed that application of (200) GA₃ induced a significantly increased in the total chlorophyll of *Solidago Canadensis*" Tara".



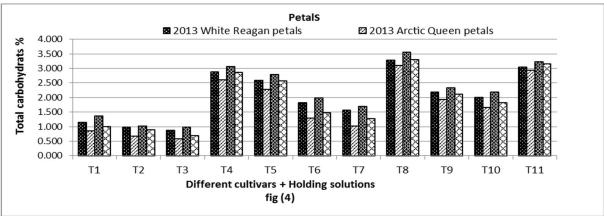


Fig. 3, 4: Effect of interaction between different cultivars, holding solution on carbohydrate (%)in petals and leaves on (*Dendesranthema grandiflorum*, Ramat.)cvs. White Reagan and Arctic Queen during seasons 2013 and 2014.

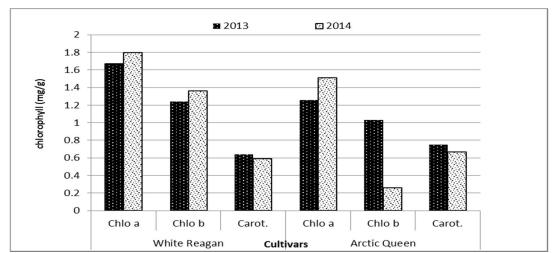


Fig. 5: Effect of different cultivarse on chlorophyll content (mg/g) in leaves of (*Dendranthema grandiflorum*, Ramat.) cvs. White Reagan and Arctic Queen during seasons 2013 and 2014.

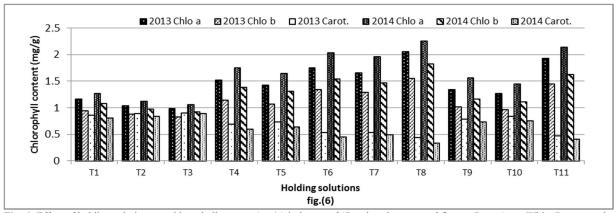


Fig. 6: Effect of holding solutions on chlorophyll content (mg/g) in leaves of (*Dendranthema grandiflorum*, Ramat) cvs White Reagan and Arctic Queen during seasons 2013 and 2014.

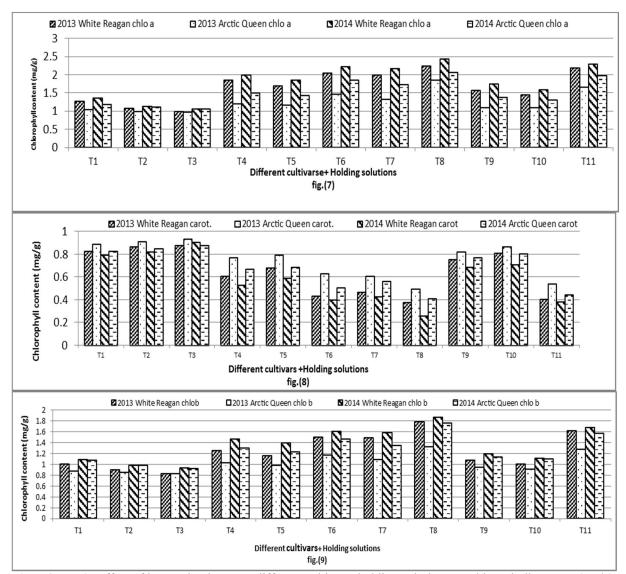


Fig.7, 8 and 9: Effect of interaction between different cultivars, holding solutions on chlorophyll content (mg/g) in leaves on (Dendesranthema grandiflorum, Ramat.)cvs. White Reagan and Arctic Queen during seasons 2013 and 2014

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      T1= Distilled water
      T2 = Sucrose (2%)

      T3= Sucrose (4%)
      T4 = Sucrose (2%) + 8-Hydroxyqunoline citrate (200 mg/l)

      T5 = Sucrose (4%) + 8-Hydroxyqunoline citrate (200 mg/l)
      T6 = Sucrose (2%) + Gibbrillic acid (0.001%)

      T7 = Sucrose (4%) + Gibbrillic acid (0.001%)
      T8= Sucrose (2%) + 8-Hydroxyqunoline citrate (200 mg/l) + Gibbrillic acid (0.001%)

      T9 = Sucrose (2%) + Citric acid (150 mg/l)
      T10 = Sucrose (4%) + Citric acid (150 mg/l)

      T11= Sucrose (4%) + 8-Hydroxyqunoline citrate (200 mg/l) + Gibbrillic acid (0.001%) + Citric acid (150 mg/l)
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